

Chapter 3

Natural-born killers: Understanding the evolutionary paths of Colombian pitvipers

Sergio D. Cubides-Cubillos, Monica M. Saldarriaga-Cordoba, Jaime A. Pereañez, Teddy Angarita-Sierra

Abstract: Historically, lanceheads, rattlesnakes, bushmasters, and hog-nosed pitvipers have received significant attention due to their medical importance and their remarkable diverse biology and venom (e.g., infra-red-sensing loreal pits, rattles, lethal toxins). Hundreds of papers and books published about pitvipers from the Western Hemisphere have established a broad and robust conceptual framework that has allowed for their understanding. However, most of these contributions have focused on temperate-zone taxa or on specific tropical countries like Brazil, Costa Rica, and Mexico. This disparity has limited our understanding of the diversity and endemism of vipers that live in geographically complex regions of tropical America, such as Colombia. Colombian pitvipers boast fascinating evolutionary histories, astonishing morphological and species diversity, and intricate geographical distribution patterns, that still await in-depth comprehension. Currently, the venom of Colombian pitvipers and its various forms hold considerable scientific importance, becoming model systems that allow for the exploration of evolutionary biology principles, from genes, individuals, and populations to macroevolutionary contexts. Based on a detailed review of available information, as well as the generation of new evidence, in this chapter we review and discuss the evolutionary lineages, venomics, and biological activities of the venoms of vipers present in Colombia.

Keywords: Diversity, fangs, venomous snakes, snakebite, tropics.



Citation: Cubides-Cubillos, SD; Saldarriaga-Cordoba, MM; Pereañez, JA; Angarita-Sierra, T. Chapter 3. Natural born killers: Understanding the evolutionary paths of the Colombian pitvipers. In Book: *Bites, venoms and venomous snakes of Colombia*; Angarita-Sierra, T., Ruiz-Gomez, FJ, Eds.; Instituto Nacional de Salud: Bogota D.C., Colombia, 2024; pp. 115–164. doi: 10.33610/866780odgbqc



Copyright: © 2024 by the authors. Open access publication under the terms and conditions of the Creative Commons Attribution (CC BY-NC-ND 4.0) license (<http://creativecommons.org/licenses/by/4.0/>).

Illustrations by:
Oscar A. Ramirez Ruiz

1. Ambush predators: Pitvipers around the world

Venom is a remarkable biological trait that has evolved independently in numerous organisms on our planet. However, this trait, which characterizes some groups of snakes, culturally generates mixed reactions and attitudes, evoking fear, fascination, and respect at the same time, making snakes stand out among all species in the animal kingdom as creatures of power [1]. Nevertheless, in the entire world only a handful of snake species exhibit exceptional venom potency or pose a significant threat owing to their sophisticated venom delivery mechanisms and specialized venom composition (see Chapter 5). Approximately 10% of all living snakes belong to the Viperidae family, encompassing vipers, adders, and pitvipers [2,3]. This family is phylogenetically divided into three subfamilies: Azemiopinae (Fea vipers), Crotalinae (pitvipers), and Viperinae (“true vipers” or “vipers without pits”).

Currently, about 383 species of viperids are recognized, grouped into 37 genera [4]. Thirteen of these genera are distributed across America: *Agkistrodon* [5], *Atropoides* [6], *Bothriechis* [7], *Bothrocophias* [8], *Bothrops* [9], *Cerrophidion* [10], *Crotalus* [11], *Lachesis* [12], *Metlapilcoatlus* [13], *Mixcoatlus* [14], *Ophryacus* [15], *Porthidium* [16], *Sistrurus* [17]; and 24 genera are distributed among the Asian, European and African continents: *Atheris* [18], *Azemiops* [19], *Bitis* [20], *Calloselasma* [21], *Causus* [22], *Cerastes* [11], *Craspedocephalus* [23], *Daboia* [20], *Deinagkistrodon* [24], *Echis* [25], *Eristicophis* [26], *Garthius* [27], *Gloydus* [28], *Hypnale* [29], *Macropviper* [30], *Montatheris* [31], *Montiviper* [32], *Ovophis* [33], *Proatheris* [31], *Protobothrops* [34], *Pseudocerastes* [35], *Trimeresurus* [36], *Tropidolaemus* [22], and *Vipera* [37].

Among all venomous snakes, the highly specialized venom delivery system of viperids is characterized by tubular front fangs enclosed in venom-conducting canals positioned on a mobile maxillary bone [38] (see Chapter 5). The specialized long teeth developed by viperids represent one of the most remarkable functional traits linked to their life history. They are primarily sedentary ambush predators that consume mostly hot-blood prey during their adulthood [38,39]. Consequently, the efficacy of this sophisticated venom release mechanism in hunting, defense, and deterrent functions has resulted in the designation of several viper genera as medically important due to the elevated annual incidence of snakebite (see Chapter 9).

In these ambush hunters, venom composition limits prey selection; hence, feeding has notorious adaptations and specificities. For example, newborn or young snakes exhibit greater lethal activity than those of their adult counterparts [40], indicating that these biochemical differences are linked with the diet changes and the evolution of the prey. Indeed, proteomic studies have revealed that ontogenetic variation in venom composition is a significant indicator for understanding toxin dynamics and actions for the two most significant Colombian pitvipers: *Bothrops atrox* [41], and *B. asper* [42,43].

To understand the evolution of snakes from a mechanical means of dominating prey (constriction) to a chemical means (venom injection [44]),

it is imperative to first comprehend the many processes through which natural selection has favored the harmony between divergence and adaptation. This balance has allowed the emergence of vast heterogeneity in the composition, function, and action of snake venom toxins. This chapter provides a brief overview of the past 50 years of research on the evolutionary and natural history of pitviper species that inhabit the tropical ecosystems of Colombia, focusing on the medically important snake species.

2. The evolutionary and geographic perspectives of pitviper colonization and diversification in the South American tropics

One of the great and pioneering contributions in the systematic and biology of pitvipers corresponds to the work of William L. Burger [45], who clarified the delimitation of several groups and the description of new characters of taxonomic use for the Viperidae family. His pioneering work suggested the division of the large *Bothrops* group into five morphologically divergent genera: *Bothriechis*, *Bothriopsis*, *Bothrops*, *Ophryacus* and *Porthidium*. Nowadays, the validity of *Bothriopsis* as a separate genus of snakes is contentious. Historically, *Bothriopsis* was considered a valid genus, but phylogenetic and genetic studies have questioned its distinction from the genus *Bothrops*. In fact, Burger's work was crucial because it triggered a series of rearrangements that allowed the description of genera derived from *Porthidium*, such as *Atropoides* [6], and *Cerrophidion* [10].

Later, Avise was one of the pioneers [46] to examine hypotheses of phylogenetic and phylogeographic inference in pitvipers from a genetic perspective used molecular markers. He studied the ancestral relationships that are unaffected by convergent selection in similar natural history traits by assessing molecular markers as independent evidence, assuming that these traits were unlikely to be influenced by the same selective pressures that act on morphological traits [47]. Since the 1990s, DNA sequences have been used to calculate the historical timelines in which different viperid clades diverged, helping to understand morphological diversification in a biogeographical context [48]. However, these initial studies examined the evolutionary value of mitochondrial genes, revealing that genes such as cytochrome *b* and NADH are the suitable predictors of phylogenetic relationships, largely due to the slower evolution of ribosomal unities [49].

Initially, Parkinson et al. [50], and Gutberlet Jr et al. [51] identified and suggested the existence of two monophyletic groups of pitvipers in America: A clade of North American species that groups *Agkistrodon*, *Atropoides*, *Cerrophidion*, *Crotalus*, *Metlapilcoatlus*, *Ophryacus* and *Sistrurus*, and a second clade that groups the other genera located in the Neotropical region. One analysis [50] recovered Americas viperids as a monophyletic group but without a suitable identification of their sister group: a single invasion of North America with a subsequent divergence between the temperate and tropical north and at least three invasions of South America by the pitviper historical ancestors. Furthermore, one

study [50] reported that the species at the time considered as *Porthidium hyoprora* corresponded to a lineage more closely related to the *Bothrops* group (coinciding with the first description [52], naming it *Bothrops hyoprora*), resulting years later in the description of a new genus, *Bothrocophias* that would include other species also included within the *Bothrops* group, such as: *B. campbelli*, *B. hyoprora*, *B. microphthalmus*, and *B. myersi* [8].

In Colombia, the significant elevation of the three Andes Cordilleras played a pivotal role in the cladogenesis of numerous pitviper species, such as the *Bothrops* species group, currently distributed allopathically [53]. The Andes Cordillera did not reach a maximum elevation of more than 40% until the Neogene era. This resulted in significant mountain formation only during the late Miocene and especially during the Pliocene, when the orogenic process accelerated quickly [54]. Hence, it is posited that the initial division among the inter-Andean pitviper groups in northern South America is attributed to a dispersal event within the Caribbean of Mesoamerica that influenced the biogeography of the region.

Molecular studies of pitvipers have primarily focused on the Neotropical region, examining various hypotheses that could explain their astonishing diversity [55–57]. Phylogenies have been used to explain the evolutionary trajectories that give origin to the types of plants and animals that currently live with us, helping to postulate theories that try to explain the diversity of organisms during geological time scales (e.g., during the Pliocene and Miocene periods) and after milestone geological events such as the Andean orogeny or the uplift of the Isthmus of Panama.

The uplift of the Andes in South America was, in fact, one of the most significant geological developments in the evolution of numerous vertebrate species [58,59]. For instance, some authors utilize historical migration analysis in populations of the lancehead (*B. asper*) to examine its recent diversification events in South America [60]. Their methodology includes the incorporation of multiple demographic processes into the inferences about patterns of divergence that are not feasible to infer or comprehend using methods based on phylogenetic trees [61,62].

Indeed, the lancehead pitviper species have demonstrated that the impact of habitat diversity is correlated with phenotypic variability, a biological phenomenon inherent to the different *Bothrops* species [63,64]. The absence of any other potential adversaries at the onset of the expansion of distinct regions in northern South America may have contributed to the spread of the lanceheads across most of the geographical regions of Colombia. Nonetheless, it is imperative to emphasize that certain genera such as *Porthidium* (*P. nasutum* and *P. lansbergii*) and *Lachesis* (*L. acrochorda* and *L. muta*) exhibit convergence with *B. asper* and *B. atrox* in some ecoregions and localities. Nevertheless, *Porthidium* species exhibit a widespread distribution in the southwestern zone of the Pacific region [65], and in the north-Caribbean region [66], being a species that are sympatric with *B. asper*.

Nowadays, the bioinformatics methods have enabled the analysis of extensive datasets, such as genomic data, allowing the evaluation of intricate historical processes with high resolution [67–69]. Furthermore, the genome has proven to be especially useful due to the accuracy and precision with which the different parameters are estimated [70–72]. In summary, systematic deficiencies in certain pitviper studies have been addressed by utilizing integrative taxonomy and various lines of evidence for species delimitation, including morphological characteristics, genetic distance analysis, molecular phylogeny, and coalescent species delimitations that can encompass geographic and ecological isolation [73–75].

2.1 Understanding how diversification events affect the diversity of pitviper species

Numerous hypotheses on Neotropical diversification have been proposed, including allopatric speciation and ecological displacement through mountain ranges [58], rivers that functioned as prezygotic barriers, or habitat fragmentation caused by climate change during the Pleistocene [76]. Continental drift of South America and the uplift of the Andes resulted in the evolutionary dynamics that underpin the foundation of diversity in numerous ecoregions of the Neotropics [77–79]. These orogenic phenomena have raised questions about how many taxonomic groups were colonized and differentiated, and which ones would later inhabit the lowlands and slopes once the Andes reached their current elevations [80]. In this way, pitviper diversity in South America is connected to biotic exchange between North and Central America throughout the Tertiary period, especially after the emergence of the Isthmus of Panama [81].

Most speciation events in tropical regions are primarily influenced by the isolation induced by landscape alteration (e.g., vicariance) that is intrinsically linked to a broader range of biological and geological processes [60]. Despite this, the current phylogeographic hypotheses proposed for South American pitviper species have not been evaluated using a consideration of the influence of demographic processes [55,57,60,82]. In fact, examining the phylogeographic patterns of distinct species with overlapping or partially overlapping ranges can reveal common events that may have a similar impact on the evolutionary patterns of numerous taxa [83]. However, inconsistencies among sympatric taxa may arise because of lineage sorting, variation in effective population size, extinction, dispersal, sympatric speciation, or non-response to vicariance events [84–86].

In contrast, the explanation for parapatric speciation along the mountainous vertical axis could be attributed to specific adaptations associated with ecological gradients [87]. Consequently, the capacity of certain species to expand into novel niches that emerged during a specific time must have been pivotal in the diversification of inter-Andean pitvipers belonging to the genera *Bothrops*, *Bothrocophias*, *Bothriechis*, *Lachesis*, and *Porthidium*. These events in northwestern South America during the early Pliocene suggest that the final uplift of the Andes could have played a significant role in the species diversification (=cladogenesis) of pitviper groups [53].

Therefore, it has been posited that the rapid increase of viperids species from a common ancestor (with heat-sensitive pits) represents a significant evolutionary milestone, enabling the successful diversification of this family of snakes [3]. From an ecological standpoint, these lineages inhabit diverse environments and habitats, ranging from tropical lowland forests in Central America and the Amazon to open areas in the Andes and in Patagonia [2,55]. However, numerous species are still poorly known with respect to their taxonomy, natural history, and phylogenetic relationships [2,28,57,88]. In fact, several groups of pitvipers show apparently endemic patterns, and because some taxa are considered species complexes, several species groups are in the process of redescription and resolution of their phylogenetic patterns (see Chapter 1).

One of the significant challenges faced by herpetology in Colombia is the limited number of molecular markers that have been examined in species of snakes. Furthermore, the limited knowledge regarding the rates of molecular evolution of various groups of snake genes precludes a comprehensive phylogeographic and historical examination of the pitvipers found in northern South America.

2.2 A family with distinct species complexes

In Colombia, as well as throughout the Andean ecosystems, the most significant environmental factors affecting the distribution of the seven known pitviper genera are elevation and orographic complexity. These factors are proposed as some of the main drivers of the phylogenetic relationships and venom diversity in several pitviper species. For instance, a select group of Colombian pitviper species attain elevations ranging from 1,500 to 3,200 meters above sea level (hereafter asl), with 1,500 m asl being the upper limit for most pitviper species inhabiting the lowlands; 3,600 m asl is the upper limit for high Andean species (such as *Bothrocophias tulitoi*, *B. myrringae*, *Bothriechis schlegelii*, and certain populations of the *B. atrox* complex) [2,75].

Within the diverse biogeographic regions of northern South American, the proportion of venomous species can range from 9% to 14% [89]. Therefore, the distributional zones of Colombian pitvipers have been primarily concentrated in areas inhabited by humans, in contrast to the notable presence of several snake populations in other forested areas [90]. As a result, the dispersion of the Colombian pitvipers is associated with primary forested areas, secondary forested areas, areas intended for agriculture at different elevations, and increasingly with urban areas (usually in expansion). Consequently, and due to their wide range of geographical distribution in Colombia, *B. asper* and *B. atrox* are the most medically important venomous snake species [64,91].

Historically, the genera *Bothrops* and *Porthidium* have been included in a substantial number of reviews and taxonomic approaches that have revealed a diversity previously unrecognized at the species level [63,92–97]. Over the past two decades, some species belonging to the

genus *Bothrops* have been the subject of numerous phylogenetic studies and biogeographic approaches pertaining to their diversification on the southern side of the western hemisphere of the earth [3,38,55,98,99]. These approaches have revealed a taxonomic complexity mainly within the *Bothrops*, although currently work is also being conducted with the *Porthidium*.

Research on the presence of cryptic diversity for several species of pitvipers found in Colombia reveals the presence of some species complexes identified through a series of common characteristics shared between several species and lineages (see Chapter 1) [53,63,75,92–95]. This difficulty in distinguishing morphologically distinct species (cryptic species) that are genetically divergent has become one of the most discussed points in work published in molecular systematics and in the process of species delimitation.

One of the objectives of this section is to document some of the pioneering research on the systematics, venoms, and evolution of Colombian pitvipers. Based on published data over the past three decades, we believe it is necessary to increase phylogeographic studies in order to analyze the cryptic diversity of recognized taxa [48,83,94,96]. This information could serve as an important tool for determining conservation efforts aimed at preserving viable populations of lineages that are yet to be discovered [83].

Below we present a description of the main groups and species complexes of the medically important pitvipers in Colombia, and we review the principal works that have laid the foundations of our knowledge. We also discuss pitviper systematics. Finally, we compile other published information about the different groups to show the association between the studies carried out on Colombian pitviper venoms and their phylogenetic correlation. Thus, we intend to show not only the phylogenetic complexity of the two main groups (*Bothrops* and *Porthidium*), but we also want to show the lineages and populations by species where there is no genetic or proteomic information.

The most medically important Colombian lancehead pitvipers: the *Bothrops asper-atrox* complex

The genus *Bothrops* exhibits more genetic divergence in comparison to the other South American pitvipers. In fact, the South American *Bothrops* exhibit paraphyletic characteristics in relation to the species found in Central America. The diversification of the genus has probably taken place in South America [60]. This hypothesis suggests that the common ancestor of all *Bothrops* was the first viperid to colonize South America sometime during the Miocene, about 10 million years ago [63]. As a result, only one lineage of *Bothrops* species (*B. asper*) spread throughout Central America, from Panama to Mexico. However, there are geographical distribution records for a second species of *Bothrops* (*B. punctatus*) in the eastern region of Panama.

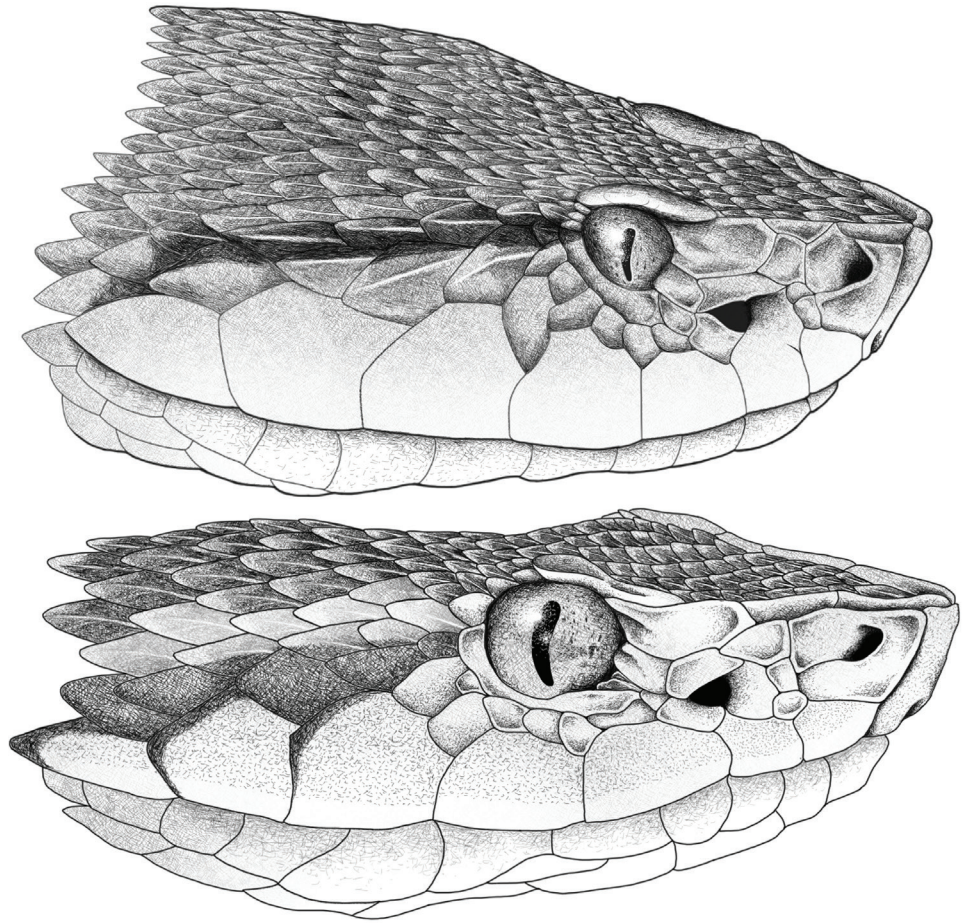


Figure 1. Head illustrations of *Bothrops asper-atrox* complex. (Top): head in lateral view of *Bothrops asper* from Melgar, Tolima, Colombia (INSZ 138). (Bottom): Head in lateral view of *Bothrops atrox* from Puerto Carreño; Vichada (BOTATR00061). Illustrations by Oscar A. Ramirez Ruiz.

Campbell and Lamar [2] suggested that Colombian populations of *B. asper* and *B. atrox* species have allopatric distributions. In fact, *B. asper* (Figure 1 Top) is distributed throughout the biogeographic regions of Choco and Magdalena Valley, with a focus on the Caribbean and inter-Andean basins [64,91], whereas *B. atrox* (Figure 1 Bottom) is found in the Orinoquia and Amazon regions. This distribution aligns with the dispersal hypothesis for the *B. atrox* species, positing that a preexisting population of forests located on the northern side of the Amazon gradually colonized the regions bordering the Amazonian River [97].

Nonetheless, there could be lineages of *B. asper* and *B. atrox* in sympatric geographical regions of Colombia's eastern and western Andes, like Boyaca, Cundinamarca, and Norte de Santander. Several studies have demonstrated that both species show recent lineage differentiation in relation to habitat diversity as well as numerous unrecognized evolutionary lineages associated with elevational gradients [53,60,92,97].

The taxonomic status of the *B. asper-atrox* complex within Colombian populations has been little explored from a genetic perspective, and there is no clear delimitation between the lineages of these species to

help recognize some of the previously described taxonomic hypotheses based on morphology. Particularly, the populations on the eastern slope of the Cordillera Oriental (*B. asper-atrox* complex) represent a challenging problem still unsolved: Some *B. atrox* mountain populations have been historically reported as *B. isabelae* [98], and *Bothrops colombiensis* [99]. *Bothrops isabelae* was initially synonymized with *B. atrox* in the 90s [33,100], but later it was recognized as a species in some of the pioneering work for South American *Bothrops* [92]. Recent analysis [101] shows *B. isabelae* together with *B. atrox* with an apparently low genetic distance. Even so, the authors recognize *B. isabelae* as a valid taxon, although a few years later it was again considered a synonym of *B. atrox* [55]. Campbell and Lamar [2] consider *B. colombiensis* as a species synonymous with the species complex formed by *B. asper* and *B. atrox* from Venezuelan populations.

Therefore, clarifying the phylogenetic connections between the Colombian *Bothrops* species and the populations of Venezuelan species/lineages would enable us to comprehend the dispersal dynamics of numerous of these widely distributed taxa, providing an alternative to the utilization of the unified species concept widely applied in analyses of geographic lineages [102]. The complexity of understanding the systematics of the *B. atrox* group is challenging [63,88,103]. Some studies suggest that the clade composed of *B. leucurus* and *B. moojeni* is the sister group of *B. atrox*; other studies indicate that *B. atrox* has closer evolutionary relationships with *B. asper* and *B. isabelae* [6,94,101,104]. Nonetheless, the *B. atrox* group originating from Colombia and Brazil requires a detailed taxonomic review, as the taxonomic hypotheses based on morphology do not align with the described mitochondrial lineages. Furthermore, these mitochondrial lineages also do not correspond with the taxonomically recognized species [103].

An example of this marked taxonomic complexity for *Bothrops* in Colombia was the naming of new species derived from *B. asper* [105]. That research provided a study that reevaluated the taxonomical boundaries within *B. asper* populations in Colombia and proposed *B. ayerbeii* and *B. rhombeatus* as new species within the *B. asper* complex. Nonetheless, the unsuitable quantity and quality of characteristics employed in the species delimitation, the ambiguous descriptions of morphologic traits and diagnostic characters [106], and the lack of a phylogenetic analysis, impedes a clear and objective distinction between the species proposed (see Chapter 1). In fact, in the initial description made by Garcia [107] no type specimen was assigned nor was the description of a new taxon made, turning *B. rhombeatus* into a *nomen dubium* (see Chapter 1).

Another crucial point mentioned by Ramirez-Chaves and Solari [106] refers that Folleco-Fernandez [105] did not describe a neotype for *B. ayerbeii* and *B. rhombeatus* nor were data on the geographical distributions provided for possible hybridization between the species nor was the relevance of distinguishing both taxa as valid species discussed. Although other authors have incorporated molecular markers to assess the phylo-

genetic relationships within the *B. asper-atrox* species complex [53]. Currently, the sequences from *B. ayerbeii* and *B. rhombeatus* populations distributed in the southwest of Colombia (Cauca and Nariño departments), proposed by Folleco-Fernandez [105], are not available in official gene repositories, being lineages synonymous with *Bothrops asper* and *Bothrocophias colombianus*, respectively. So, further studies should include a larger or sufficient series of individuals collected in different habitats and analyzed with an integrative taxonomic approach in their sampling methodology [60,97].

In order to highlight the absence of genetic and venomous information of some representative clades and lineages within the Colombian pitviper species, we present as a novelty a heat map that shows the phylogenetic relationships of the *B. asper-atrox* complex and some proteome data published for some species in the last two decades. Thus, Figure 2 shows the result of a molecular phylogeny constructed from fragments of two mitochondrial genes, and supported by some genera of pitvipers as out groups (Appendix). We have recovered a topology that includes terminals for different populations and lineages, as well as the recently described Colombian *Bothrocophias* species [75]. We used three terminals from three populations in Brazil, including two terminals reported as species but recently synonymized for the species *B. atrox*. We used data from eight groups of protein/toxin families that have been frequently reported in venomous studies, and we performed the association of the percentages of these main components to present hypotheses related to the conformation of the clades and their geographical distribution (Figure 2).

We suggest that *B. asper* from Colombia presents three large and well-supported phylogeographic lineages that are morphologically and ecologically diverse with genetic divergences between 2.5% and 4%. On the other hand, *B. atrox* presents undefined population groups whose phylogenetic relationships still need to be well established (populations west of the Cordillera Oriental-Cundinamarca vs. populations east of the Cordillera Oriental-Meta, Vaupes and Caqueta). Furthermore, the geographical and taxonomic limits with its synonymous species/lineage *B. isabelae* (also distributed in the eastern versant of the Cordillera Oriental in some parts of Venezuela) continue to be a taxonomic problem to be resolved. Here we use information from ongoing work to indicate that revision of the taxonomic status of the species/lineage *B. isabelae* could be reported later.

It is important to note that the sister genus of *Bothrops*, *Bothrocophias*, has only three species with genetic data and a single published proteome (see section 3 in this chapter). Likewise, we were not able to calculate this relationship analysis for the proteomes of the *B. ayerbeii* and *B. rhombeatus* lineages due to the absence of genetic data available. Finally, the absence of venomous data for two important lineages within *B. asper* (populations from the Caribbean region and Middle Magdalena) is noted here.

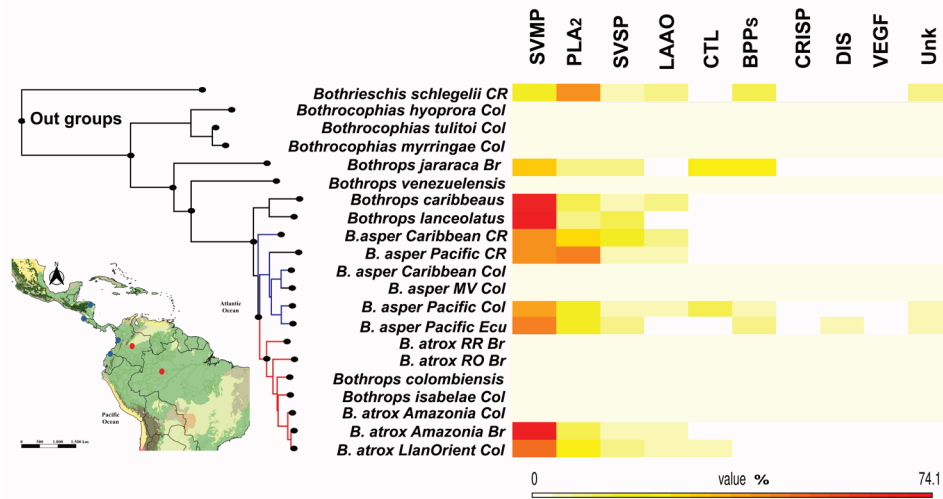


Figure 2. Molecular phylogeny of concatenated genes (MT-ND4 and MT-CYB) sequences, produced by [3,50,53,63,75,94,96], and Cubides-Cubillos et al. (unpublished data) using maximum likelihood (ML) and a proteome heatmap of the Colombian pitvipers (*Bothrops asper-atrox* complex). Phylogenetic topology retrieves the species and their most important lineages and those for which genetic and proteomic information is available (see bibliographic information in Section 3). Besides, the nodes that support branches with bs > 90% are shown. Abbreviations: lineages/populations (Caribbean and Pacific Col: Colombia; Pacific Ecu: Ecuador; MV Col: Magdalena Valley, Colombia; RR Br: Roraima region from Brazil; RO Br: Rondonia region from Brazil; Amazonia Br: Amazonian region from Brazil; Amazonia Col: Amazonian region from Colombia; LlanOrient Col: Orinoquia region from Colombia; and protein family names: SVMPs: metalloproteinase; PLA₂s: phospholipase A₂; SVSPs: serine proteinase; LAO: L-amino acid oxidase; CTL: C-type lectin/lectin-like; BPP: bradykinin-potentiating peptide; CRISP: cysteine-rich secretory protein; Unk: nucleotidase; phosphodiesterase; hyaluronidase; nerve growth factor; peptides and/or non-protein compounds.

With relation to the species *B. atrox* and the various lineages found in previous phylogenetic analyses, only records of proteomic data were found for a population from Colombia (Llanos Orientales region, Meta department) and the northern region of Brazil (Amazonas State). It remains under consideration for subsequent analysis, whether the populations of *B. atrox* distributed along the eastern versant of the Cordillera Oriental (encompassing Boyaca department), populations of the Colombian Amazon in the foothill region (Caqueta and Putumayo departments) of the Eastern Andes and those from the Amazon lowlands (departments of Guainia, Guaviare, Vaupes, and Amazonas), would require a review of their venoms, as well as other molecular data. Likewise, data from highland populations on the eastern versant of the Cordillera Oriental identified as the *B. isabelae* lineage, could show certain types of venom adaptations such as biochemical diversity, functional specialization, and evolutionary dynamics as gene duplications, diversification and rapid evolution of toxin genes, which represents a challenge in future research for mountain populations pitvipers genus *Bothrops*.

The hemorrhagic potency of several types of Snake Venom Metalloproteinases (SVMP) is one of the important factors in the biological action of *Bothrops* venoms. Its relative abundance in venoms is related to local and systemic hemorrhage [43]. Our relationship analysis showed that the species and populations with the highest content of SVMPs were geographically distributed in both dry and humid tropical regions. Furthermore, venom composition exhibited high variability within closely related lineages, indicating a possible ecological response. Probably, the heterogeneous distribution and dispersal of species/lineages is a possible cause of the synergistic effects between several types of SVMPs. In fact, hemorrhage, which also contributes to myonecrosis [108], is one of the most documented effects in snakebite accidents of all members of the *B. asper-atrox* species group.

The PLA₂, the second most abundant toxin in the *B. asper-atrox* species venom is responsible for myonecrosis and is closely related to the

digestive functions of the venom [109-110] (see Chapter 5). This toxin usually represents about 20-40% of the proteome venom composition of the *B. asper-atrox* species group but shows moderate variability. In fact, the variability in PLA₂ percentage seems not to be related to a specific clade, region, or climate. However, due to large information gaps, this pattern could change in future studies or could be associated with ecological variables such as prey availability, habitat loss, or anthropogenic pressure.

Enzymes incompletely mimic the action of thrombin, and Snake Venom Serine Protease (SVSPs) exert a variety of actions on hemostasis and the kinin system or kinin-kallikrein system [111,112]. They constitute the third most important group of toxins and, in fact, show minor variation within the species/lineages of the *B. asper-atrox* group. Reports of snakebite accidents do not place much emphasis on variations in coagulopathies or inflammation, as these are typical symptoms of bothropic envenomation (see Chapter 9). Evolutionarily, this appears to be a characteristic intrinsically linked to constructive interactions with other components of the venom (see Chapter 5 and 9).

The Colombian hognose pitviper: *Porthidium* species group

Hognose pitvipers inhabit Central and South America, and it is hypothesized that their dispersal occurred along the Pacific coast during the final phase of the emergence of the Panamanian isthmus (3.5 mya), even after its formation [63]. This dispersion might have resulted from interspecific competition with other viper species such as *B. asper*, a species that has been identified as another pitviper that colonized South America before the Panamanian Isthmus [2,53,63]. The species of *Porthidium* are short-bodied terrestrial snakes that do not exceed 100 cm in length and whose hypothetical evolutionary origin is related to Central America [2,3,13,14,113].

This genus shares cryptic dorsal color patterns and a clearly defined canthus rostralis (rostral upward scale; see Figure 3) that presumably denotes important adaptations for the hunting forms typical of ambush predators [2]. In the 1990s, phylogenetic relationships between different species were initially reviewed using morphological characters and molecular data [6,50,104].

Currently, various taxonomic studies have revealed the evolutionary and systematic relationships between the different species that support the taxonomic validity of nine species of the genus *Porthidium*: *P. dunni*, *P. hespere*, *P. ophryomegas*, *P. volnicacum*, and *P. yucatanicum* with a strong affinity towards arid zones; and *P. nasutum*, *P. porrasi*, *P. lansbergii*, and *P. arcosae* that inhabit lowland tropical rainforests [95]. It is important to highlight that the description of the latest species for Central America, *P. porrasi* [114], was based on an analysis of adjacent populations of *P. nasutum* from Costa Rica. This implied the initiation of a discussion on phylogenetic diversity within the genus and marked the first strong revision suggested for northern South American populations [95].

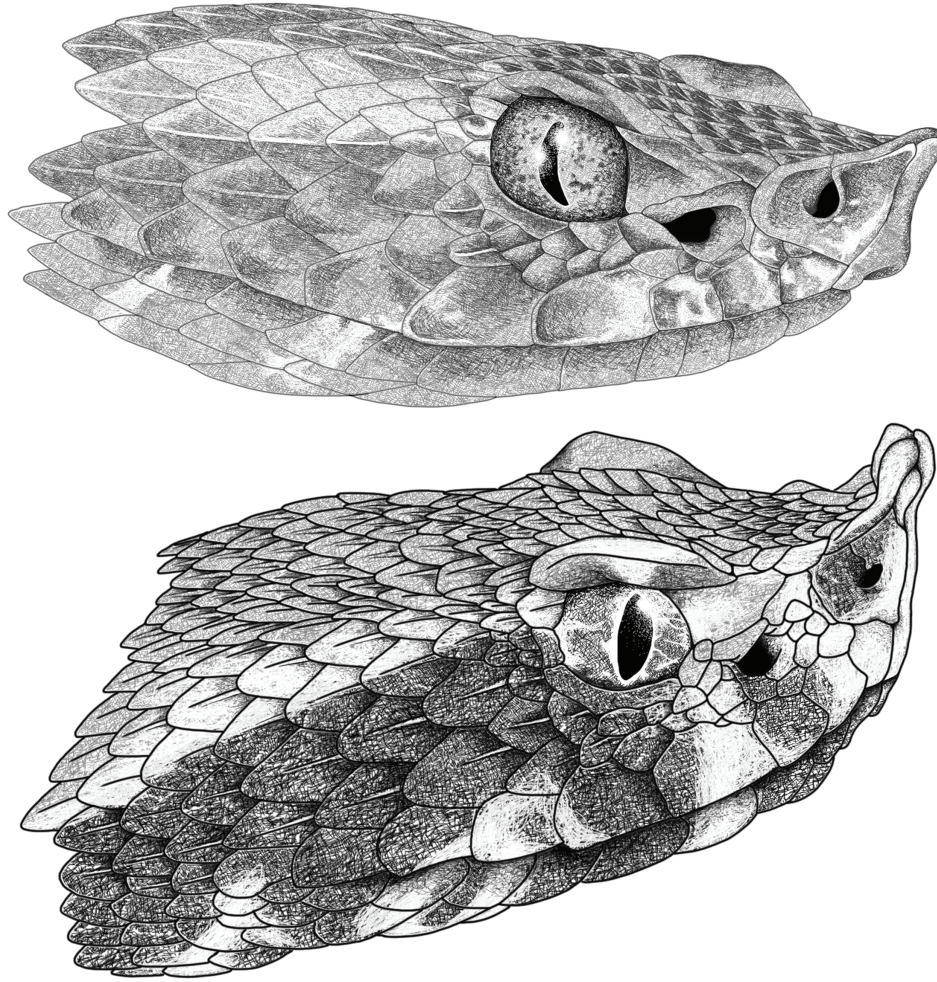


Figure 3. Head illustrations of *Porthidium lansbergii-nasutum* species complex. (Top): Head in lateral view of *Porthidium lansbergii* (INSV-SR-89) from Yondó, Antioquia, Colombia. (Bottom): Head in lateral view of *Porthidium nasutum* (QCAZR 15395) from Tundaloma Lodge, Durango, Ecuador. Illustrations by Oscar A. Ramirez Ruiz.

In Colombia, *P. nasutum* [115], and *P. lansbergii* [116] are closely related species, characterized by cryptic morphological traits such as the ‘hog-nose snout’ character. *Porthidium nasutum* is distributed from Mexico through the Isthmian-Atlantic moist forests, transitioning into the Choco-Darien ecoregion, as well as the biogeographic Chocoan ecoregion, including Ecuador [2]. Meanwhile, *P. lansbergii*, is distributed in the Isthmian-Pacific dry forests of Panama and the Colombian Caribbean, before reaching the tropical dry forest in northern Venezuela and the Choco-Darien region, where it transitions into the Uraba humid forests and the evergreen and dry forest in the Magdalena Valley [2,117].

Most of the previous scientific contributions for *P. nasutum* and *P. lansbergii*, performed phylogenetic approaches based on geographically restricted and small individual sampling [117,118]. The taxonomic complexity observed in Panama and in the diverse Colombian populations warrants large-scale research. For example, Castoe et al. [95] demonstrated that *P. nasutum* is a paraphyletic clade that shows two genetically well-differentiated lineages in Costa Rica and Ecuador.

Similarly, in our phylogeny of *P. nasutum* we recovered three principal lineages: Clade 1 and Clade 2 from Costa Rica and Ecuador, matching with previous results reported Castoe et al. [95], while Clade 3 clustering specimens distributed from the northeast of the Colombian Andes, and those from the central-eastern region of Antioquia department. Yet, in our phylogenetic analysis, the Colombian population (central-eastern Antioquia) was recovered as a clade related with the different lineages of *P. lansbergii* group. Previously, Cisneros-Heredia and Yanez-Muñoz [119] suggested that the Ecuadorian population ought to be regarded as a novel species, distinct from *P. nasutum* in Central America. However, our phylogeny does not include samples from the Chocóan populations of *P. nasutum* (Colombian Pacific Coast) or from the upper Magdalena River basin (Huila department), which could limit the resolution of the phylogenetic relationships between the *P. lansbergii* and *P. nasutum* Colombian lineages, as well as the inferences derived from this.

Our molecular phylogeny highlights that *Porthidium lansbergii* exhibits a wide range of diversification across diverse ecoregions encompassing elevation ranges from sea level to 1,200 meters. The Colombian lineages exhibit paraphyletic groups: Populations from the Darien-Panama are distinguished from those located in the Colombian Caribbean ecoregion, the Magdalena River basin, insular populations, and those to the east in Venezuela. Furthermore, certain genetic sequences associated with the two subspecies of *P. lansbergii* (*P. l. hutmanni* and *P. l. rozei*) were grouped in a polytomic clade, indicating a close association between the lineages of the Caribbean coast and Magdalena Valley with individuals analyzed from Venezuela, as also reported by De Arco-Rodríguez et al. [118].

Only a few species of *Porthidium* have published proteome composition. Despite of *Porthidium* second genus of viperid with the greatest medical importance in Colombia (see Chapter 9), only one population *P. lansbergii* has a proteome description

Nevertheless, the relationship analysis of molecular phylogeny and proteomic data of *Porthidium* species shows that there are significant differences between the proteomes of sister species. However, due to the fragmentary state of knowledge about their venom compositions, it is not possible to propose an explanation for the apparent variability observed between the lineages, nor about the concordance between phylogeographic lineages and ecoregions, or ecological characteristics.

The main toxins that vary between species correspond to the three main toxin groups: SVMP, PLA₂, and SVSP. Interestingly, these toxins vary in clades distributed in divergent climatic regions. However, missing data for several populations/lineages within the species *P. nasutum* and *P. lansbergii* could indicate different adaptive processes between populations of the same species. Therefore, we encourage both Colombian and foreign researchers to tackle biogeographic hypotheses that attempt to explain how the Andes promotes the random explosion of phenotypes (morphology, venom compositions, color patterns, etc.), as well as how selective

pressure on venoms correlates or does not correlate with lineages and proteome variability [188]. These hypotheses and speculations require future studies that significantly reduce the uncertainty surrounding the evolution of lineages within the *Porthidium* genus and their relationship with the functional response of their venoms.

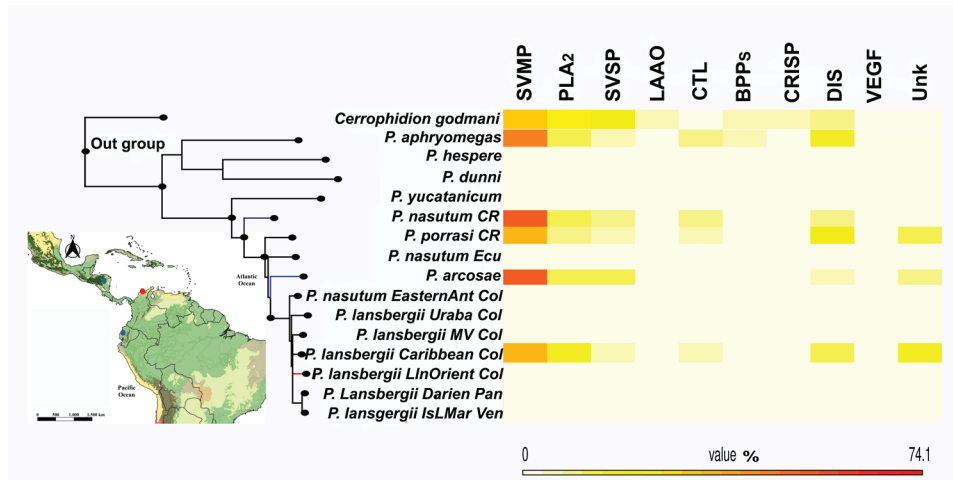
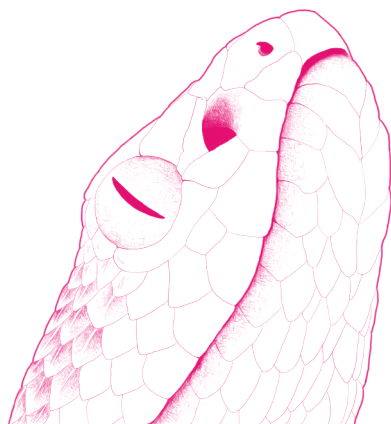


Figure 4. Molecular phylogeny of concatenated genes (MT-ND4 and MT-CYB) sequences, produced by [95, 114], and Cubides-Cubillos et al. (unpublished data) using maximum likelihood (ML), and proteome heatmap of the Colombian pitvipers (*Porthidium* genus). Phylogenetic topology retrieves the species and their most important lineages, and those for which genetic and proteomic information is available (see bibliographic information in Section 3). Additionally, the nodes that support branches with BS > 90% are shown. Abbreviations: lineages/populations (CR: Costa Rica; Ecu: Ecuador; EasternAntCol: Eastern of Antioquia department, Colombia; Uraba Col: Uraba region from Colombia; MV Col: Magdalena Valley, Colombia; Caribbean Col: Caribbean region from Colombia; LlanOrient Col: Orinoquia region from Colombia; Darien Pan: Darien region from Panama; IslMarg Ven: Margarita Island from Venezuela); and protein family names: (SVMPs: metalloproteinase; PLA₂s: phospholipase A₂; SVSPs: serine proteinase; LAAO: L-amino acid oxidase; CTL: C-type lectin/lectin-like; BPP: bradykinin-potentiating peptide; CRISP: cysteine-rich secretory protein; Unk: nucleotidase; phosphodiesterase; hyaluronidase; nerve growth factor; peptides and/or non-protein compounds).

Giant Colombian pitvipers: The South American rattlesnakes and bushmasters

The South American rattlesnake *Crotalus durissus* (*sensu lato*) is a remarkable species of *Crotalus*; its venom is of significant medical concern due to its characteristic actions of neuromuscular paralysis, rhabdomyolysis, acute kidney injury, and coagulopathy [120] (see Chapter 5 and 9). Recent research documents the diversity of subspecies for the species *C. durissus* [121], reported as follow: *C. d. durissus* [11]; *C. d. cascavella* [9]; *C. d. collilineatus* [52]; *C. d. cumanensis* [122]; *C. d. marajoensis* [123]; *C. d. maricelae* [124]; *C. d. ruruima* [123]; *C. d. terrificuss* [37]; and *C. d. trigonicus* [125]. These authors include two additional species from northern South America, *C. unicolor* [126] with a geographic distribution restricted to Aruba and *C. vegrandis* [127] with a geographic distribution restricted to Venezuela and previously regarded as a subspecies of *C. durissus* (see Figure 5, Top).



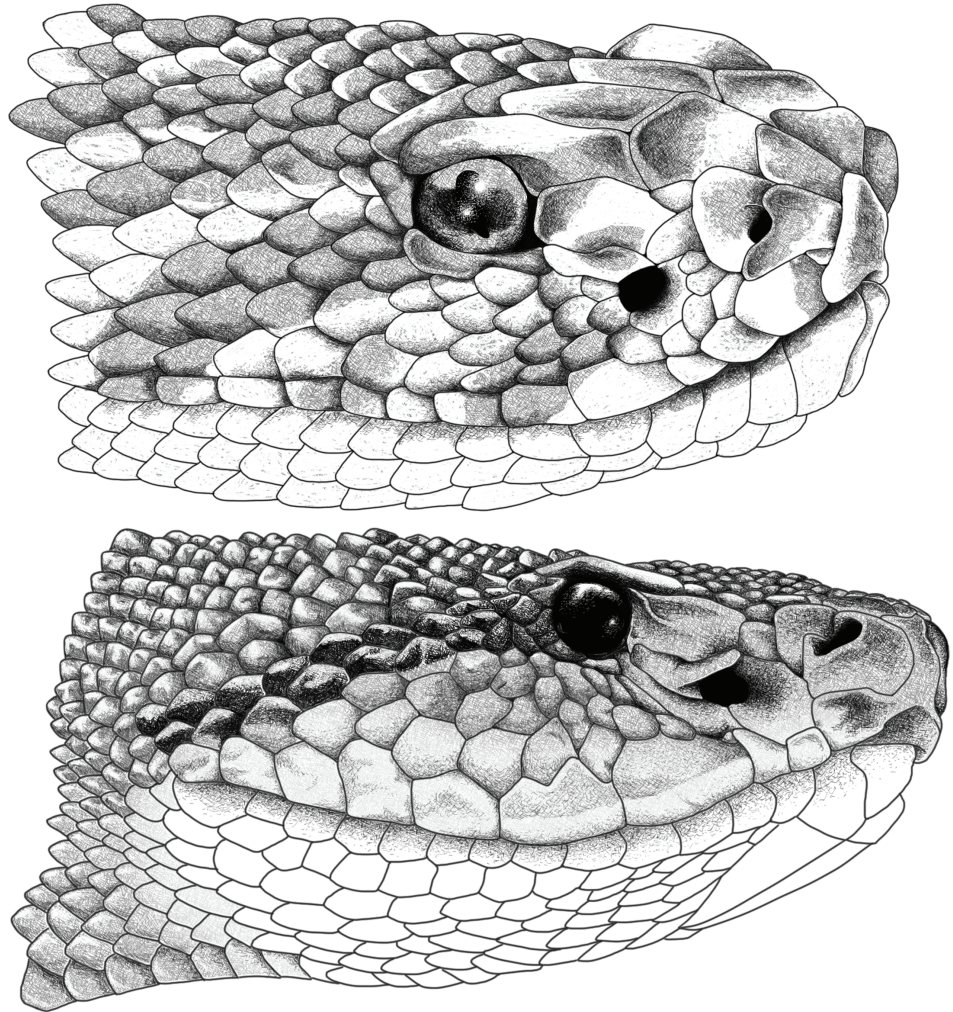


Figure 5. Head illustrations of giant Colombian pitvipers. (Top) Head in lateral view of *Crotalus durissus* from the municipality of El Paso, Cesar Colombia (INSZ 115). (Bottom) Head in lateral view of *Lachesis muta* from Mirití-Paraná, municipality of La Pedrera, Amazonas, Colombia (IAvH-R-8721). Illustrations by Oscar A. Ramirez Ruiz.

Nonetheless, certain taxonomists have begun to recognize the subspecies as distinct species [128,129]. In recent work, the *C. durissus* group (including *C. vegrandis*) has been phylogenetically recovered as a sister group to *C. simus* (Central American rattlesnake) [121]. However, it appears that the cryptic diversity in this group is the result of processes of isolation. Furthermore, the molecular phylogeny published for the *C. durissus* group does not consider individuals from the trans-Andean and cis-Andean regions of Colombia.

In Colombia, *Crotalus durissus* is distributed in the trans-Andean region of the country over the lowlands of the Caribbean region, extending into the Magdalena Valley, with disjunct populations in the middle and upper Magdalena River basin. In the cis-Andean region, it is found in the well-drained high plains of the Orinoquia [2]. It is regarded as a species of terrestrial habits and is a prevalent resident of open areas that are relatively dry [130]. This rattlesnake exhibits ecological flexibility, allowing it to be active both within forests and outside their edges, and it is commonly found in heavily human-altered open areas [130,131]. Though it is one of the most venomous species in Colombia and one of the most

illegally traded [89], information on its natural history, registers of its occurrence, and evaluation of its state of conservation are still missing [64] (see Chapter 1). The disjunct distribution pattern of its populations within the trans-Andean region, as well as between the cis-Andean and trans-Andean populations, suggests that it is very likely that *Crotalus durissus* is composed of more than one evolutionary lineage that has diverged due to geographic and ecological isolation. Recently, significant intraspecific differences were found in the composition and biological activities of *C. durissus* venom among populations distributed in the middle Magdalena River basin, the Llanos Orientales foothills, and the Colombian Caribbean coast (see Chapter 5). These results suggest that the hypothesis that *Crotalus durissus* is a species complex with multiple evolutionary lineages that have diverged due to geographic and ecological isolation is plausible. However, future phylogenetic studies, including representative samples from all Colombian populations, as well as populations distributed in Brazilian ecosystems, are expected to help resolve these uncertainties.

The snakes of the genus *Lachesis* [12], typically called “bushmaster”, are the largest pitviper snakes so far known, reaching more than 3.5 meters in length [2,89]. The snakes of this genus are the only oviparous species among all American pitvipers [132]. The species of the *Lachesis* genus can be divided into Central American and South American lineages, with an estimated divergence between 6 and 18 million years ago. The divergence within the Central American species seems to have occurred between 4 and 11 million years ago, likely due to the uplift of the Talamanca Range. However, the differentiation within South American lineages occurred only about 800,000 to 300,000 years ago [48,133]. Specifically, the emergence and structuring of the three Andean mountain ranges in Colombia has been proposed as the mechanism of isolation between *L. muta* (restricted to the Amazon) and the Central American bushmaster species (*L. melanocephala* and *L. stenophrys*), and *L. acrochorda*, the only *Lachesis* species inhabiting the trans-Andean region, distributed in the humid forests of the South American Pacific region, the Darien region in Panama, and the middle Magdalena River basin.

Recent literature reviews for *Lachesis* document the rarity of Colombian bushmasters, the number of individuals (specimens housed in biological museums; see Chapter 1), a few facts about the nature of the genus, and the small number of snakebite accidents annually [134]. In fact, it is surprising, given that *Lachesis* is considered the largest venomous snake species in South America, reaching up to 350 cm in length [89], and with a distribution encompassing four of the five Colombian ecoregions.

The systematics and taxonomy of the *Lachesis* species are the best studied aspects of the genus. In fact, this research has explored hemipenial morphology, color pattern and scale counts. Despite of these Colombian populations of the two species distributed in the country are poorly represented in these studies or have not been included [134].

It is important to highlight that in Colombia, the bushmaster species *L. acrochorda* (see Figure 5, Bottom) and *L. muta* have allopatric distribu-

tions: Populations of *L. acrochorda* are distributed in the Choco-Magdalena biogeographical province in western Colombia, and populations of *L. muta* are distributed on the eastern slopes of the Cordillera Oriental and in the Amazon ecoregion [135]. Colombian populations of bushmaster species have been poorly represented in phylogenetic assessments. For example, evolutionary relationships between the Colombian populations of *L. muta* in the Amazon and the populations of *L. muta* in the Atlantic Forest of Brazil have not been discussed.

Knowledge of the venom and toxicological characteristics of Colombian bushmasters is still fragmentary. The protein composition of *L. acrochorda* venom is known from populations from the Pacific Coast and middle Magdalena River basin in Colombia, allowing the appreciation that the conformation of *Lachesis* venom is apparently very conservative in all species of the genus [136,137]. However, in the future some studies of the molecular systematics and new venom analyses will be published among populations of *L. acrochorda* from the southwest, northeast, and Pacific coast of Colombia (A.M. Franco-Vasquez personal communication 2024). For Amazonian *L. muta* from Colombia few records are available; and therefore, no biological and natural history information has been documented [138]. Thus, given medical importance of bushmaster populations from the Colombian Amazon, it is a priority to conduct venom proteomic characterization that allows to compare the venom compositions between populations from Colombia and Brazil.

Perspectives on poorly known species of Colombian pitvipers

The last 20 years of herpetological research have allowed us to clarify several of the different biological processes for the main species of pitvipers in Colombia. *Bothrocophias* and *Bothriechis* represent one of the challenges for researchers working with genetic and molecular data. The recent description of two new species of *Bothrocophias* reveals the importance of evaluating the cryptic populations of some species with a wide distribution (like *B. microphthalmus*) and of a genetic approach to some groups of species present in several South American countries [75].

Similarly, the proposal of new species *Bothriechis* for Colombia opens a space for deeper research on the venoms of the only group of arboreal pitvipers in Colombia [139]. Nevertheless, these efforts were hindered due to deep inconsistencies in species delimitation within Colombian populations, as well as the feeble evidence and unsupported new lineages proposed by Arteaga et al. [139] (see Chapter 1). Recently, Reyes-Velasco [222] re-evaluated the taxonomic proposal by Arteaga et al. [139], demonstrating that these authors misinterpreted their results. The genetic differences they interpreted as species boundaries may instead reflect clinal variation, not independent lineages. The rest of the taxa proposed by Arteaga et al. [139], including *B. khwargi*, *B. klebbai*, *B. rahimi*, *B. rasi-kusumorum*, and *B. torvus*, do not hold up under more rigorous analysis and should be synonymized with *B. schlegelii*. Therefore, we agree with Reyes-Velasco's [222] conclusions and recommend that future publica-

tions dealing with Colombian populations of *Bothriechis schlegelii* refer to it as the *B. schlegelii* species complex (see Chapter 1).

Published and available molecular data for different pitviper species inhabiting in Colombia were summarized in Table 1. However, despite several main phylogenies published on Neotropical pitvipers that provide our current understanding of the evolutionary paths of Colombian pitvipers, several taxa and populations of pitvipers in Colombian ecoregions are poorly represented or were not included in these phylogenies [3,55,63,92,93,101,55]. Therefore, evolutionary paths of Colombian pitvipers still fragmentary.

Bothrops asper has been the only Colombian species extensively genetically tested from molecular supports using partial sequences of the cytochrome b and NDH4 genes [53,60]. Also, parallel studies at the proteomic level could reveal the presence of geographical lineages for the main groups of species that are distributed North of South America, given that new research is being developed in the field of molecular systematics. We expect that the results of ongoing projects investigating the evolutionary trajectories of *B. atrox*, *P. nasutum*, *P. lansbergii*, and *Crotalus durissus* will arise in the coming years (Cubides-Cubillos et al.; and Arias-Sosa et al. in press), helping to unveil the complexity of the phylogenetic history of these Colombian pitvipers.

Table 1. Molecular marker data published for pitvipers from Colombia and other countries in the neotropical region.

Species	Cytochrome b	NDH4	Nuclear genes	Microsatellites
<i>Bothriechis schlegelii</i>	+*	*	-	-
<i>Bothrocophias hyoprora</i>	+*	+*	-	-
<i>Bothrocophias tulitoi</i>	+	+	-	-
<i>Bothrocophias myrringae</i>	+	+	-	-
<i>Bothrocophias myersi</i>	-	-	-	-
<i>Bothrocophias colombianus</i>	-	-	-	-
<i>Bothrops asper</i>	+*	+*	*	-
Lineage <i>B. rhombeatus</i>	-	-	-	-
Lineage <i>B. ayerbei</i>	-	-	-	-
<i>Bothrops atrox</i>	+*	+*	+*	*
<i>Bothrops bilineatus</i>	+*	+*	*	-
<i>Bothrops punctatus</i>	-	-	-	-
<i>Bothrops pulcher</i>	-	-	-	-

<i>Crotalus durissus</i>	+*	+*	+*	-
<i>Lachesis acrochorda</i>	+*	+*	-	-
<i>Lachesis muta</i>	+*	+*	-	-
<i>Porthidium lansbergii</i>	+*	+*	*	-
<i>Porthidium nasutum</i>	+*	+*	*	-

(+) Data for Colombian species. (*) Data for species in other countries. (-) unpublished data. For the species *P. lansbergii* and *P. nasutum*, the molecular data correspond to research in progress developed by Cubides-Cubillos et al. as well as the data for *C. durissus* corresponding to a review developed by Mario Vargas-Ramirez et al.; the other sequences were developed by [3,50,53,63,75,95,96,104], and [114].

3. Biological activities and proteome of the Colombian pitviper venoms

The evolutionary selection of specific types of toxic components appears to have been somewhat limited. Therefore, it is posited that the venomous proteins belong to a limited number of protein families [140]. Indeed, the restricted variety of proteins has undergone rapid evolution in its initial location, resulting in the production of numerous biological functions [141]. Hence, as venoms are trophic adaptations that facilitate subduing prey and their effects are highly variable according to the type of prey and its availability, the diverse toxic secretions of snakes exhibit significant variation in their composition [142].

Venom composition in snakes exhibits variation across all taxonomic levels (between families, genera, and species), yet understanding its mechanisms remains a subject of research [143,144]. A plausible explanation for this variation may require the conduct of adaptation evolutionary studies, coevolution, and the establishment of a phylogenetic and ecological framework [110,145,146,144]. The coarse description of the variability in venom composition has an impact on applied and biomedical sciences, influencing the efficacy of antivenom therapies and the treatment of snakebites, as well as the development of new drugs [111,147] (see Chapter 6 and 10).

Several countries in South America exhibit a high prevalence of bothropic snakebites associated with serious and life-threatening envenoming (see Chapter 9). This has led to the stigmatization of the remaining pitviper species, as well as similar non-venomous snakes, resulting in their deliberate killing by diverse human communities across their distribution ranges. Nonetheless, apprehension regarding pitviper species belonging to the genus *Bothrops* is not unfounded. Indeed, due to their capacity for inhabiting diverse ecological zones, *Bothrops* species are among the most dangerous venomous snakes in tropical regions of Central and South America. Particularly in Colombia, bothropic envenoming accounts for 62% of annual cases [111,148] (see Chapter 9).

Current research, focused on the study of venoms employing a multidisciplinary approach, using “omics” technologies such as genomics, transcriptomics, proteomics, and, more recently, metabolomics is used to investigate

low molecular weight components [149] of venoms. These research fields have had a significant impact on current biomedical science [150]. Hence, by incorporating advances in high-resolution mass spectrometry (MS) [151], and de novo and database-dependent annotation methods that need minimal amounts of venom [152], enables identification of toxin families, individual toxins, unique isoforms, splice variants, amines, amino acids, and alkaloids.

More than a hundred Neotropical pitvipers are recognized worldwide [4]; and given their extensive distribution, they constitute a medically important event with a significant impact on health, since they cause more than 2.7 million snakebites per year [108]. However, snake venoms, at the same time, are a source of raw materials for the acquisition of new molecular compounds [153], which facilitate various biotechnological advancements in molecular biology and biomedicine (see Chapter 10).

Chippaux et al. [154] paved the way for the study of snake venom, showing that these multifaceted secretions can differ at different levels and are affected by various intrinsic and extreme factors [144]. Currently, studies in venomous biology serve as models for studies in evolutionary genetics [155,156]. In the following sections, we will provide a summary of the knowledge about venomous of Colombian pitvipers. We will also discuss the gaps and research perspectives achievable in the near future.

3.1 The complexity of venom: more than a hunting weapon

Pitviper venom contains a complex mixture of proteins and enzymes with different biological activities [157]. Their primary function is to immobilize, pre-digest, and kill the prey (see Chapter 5). This enabled the transition from a mechanical subduction mechanism (body pinning and constriction) to a chemical subduction mechanism (the use of venom) [158]. Thus, the venom system became a main feature of advanced snakes [159], and it is an important trait for understanding the evolution and ecology of various snake species [141].

These complex mixtures of proteins are produced by specialized buccal glands that are homologous to the Duvernoy and salivary glands and are also present in other members of the Colubroidea superfamily [157,160]. These toxins originated from duplication events of genes encoding proteins with normal physiological functions (whose resulting proteins are recruited by the venom gland) and that show a selective expression on the structure [141,161].

Over time, venom gland-encoded proteins were subjected to selection pressures that resulted in their structural and functional modification and even suppression of their expression in the gland [161]. However, information about changes in the structural components and expression, as well as gene duplication related to the origin of the actual functions of various proteins and toxins in general, is still not well understood. Thus, understanding the order of such events that explain the origin of any protein is a challenge yet to be resolved [162].

Snake venoms present great intraspecific and interspecific variability, largely because these are ecological traits that evolve dynamically. There are several factors that influence their synthesis such as seasonality, geographic distribution, ecological variation between populations, and the variety in the diet that intrinsically diverges with the age/sex of the individual [154,163,164]. Since the properties of these venoms are useful when defending against predators, human beings have become part of this interaction.

For example, venoms from three clades of spitting cobras have the ability to activate mammalian sensory neurons, inducing pain [165]. In fact, the divergence of the spitting cobra clades in Africa (6.7 Mya) could be associated with the evolution of bipedalism and the development of larger brains in hominids [165], indicating that “spit venom” could be a mechanism that could have evolved in response to interactions with ancient hominins. In this sense, snakes will need to produce copious amounts of venom, which would implicitly make it difficult to infer whether the venom has a high-energy cost when compared to other ecological traits. In fact, it is difficult to determine experimentally this earlier premise due to the difficulty of experiments that would require specific prey management, taxon-specific toxicity testing, and, furthermore, an autonomous way in which snakes can modulate venom production and injection [166].

In fact, today those circumstantial or provoked events in which a snake bites a human has been continued, technically called “snakebite accidents” [167]. Hence, this incident leads to an intoxication resulting from venom inoculation from snakebite, resulting in a clinical case that is regarded by the World Health Organization (WHO) as one of the numerous neglected tropical diseases (NTDs; see Chapter 9). Unfortunately, these snakebite cases occur more often in developing countries, where almost 95% of the total number of snakebite cases worldwide are reported [168]. Moreover, since these accidents are frequent in low-income rural communities with limited access to healthcare services, addressing these envenomations is challenging due to the limited availability of antivenoms and the high costs of transporting the patients [168,169] (see Chapter 6).

In perspective, venoms are a rapidly evolving trait; it has been inferred that environmental factors can generate adaptive pressures that modulate their functional variation across species. As an adaptive characteristic used both for feeding and for deterring or defending against potential predators, developing effective antivenoms to neutralize their envenomations is currently a significant challenge, since these therapies must account for the wide intraspecific divergence in venom composition. In fact, recent research that reports on divergence of crotoamine content in populations of the genus *Crotalus durissus* should be a warning about the need to develop an antidote capable of neutralizing this toxin [170,171].

One of the main “targets” of the venom of some snakes is the nervous system of the potential prey, even of one possible predator [172]. The presence of neurotoxins in these venoms is attributed to their capacity to disrupt the normal functioning of the central nervous and/or peripheral

nervous system [173,174]. The high specificity and selectivity of neurotoxins found in rattlesnake venoms have been widely used to study the structure-function of receptors and other elements involved in neuromuscular transmission, neurological mutations, and the causes of various neurological conditions [175] (e.g., Parkinson and Alzheimer, see Chapter 10). However, today, little research is available on the effect of toxins on the peripheral nervous system and some specific organs [172]. Researchers will continue to gain understanding about the targeted administration of drugs for the treatment of various nervous conditions, tumors, and neurodegenerative diseases in the future. In the interim, the persistent pursuit of research aimed at identifying novel ophidian peptides and comprehending their structure-function correlation with cellular receptors, will ultimately lead to the discovery of distinct molecules that have the potential to be utilized in diverse biomedical therapies (see Chapter 10).

In Colombia, the colonization of new niches in different mountainous, dry lowland, or humid tropical ecosystems must have played an important role in the phenotypic diversity of pitviper venoms that could be shaped by local adaptation to different habitats within ecoregions (e.g., the intraspecific differences in the venom of *Crotalus durissus* [171], see below). Nevertheless, further research efforts are still needed to understand the relationships between the composition of venoms, their ecological context, and their link to habitat transformations caused by human activities. For this, it is necessary to develop a more robust conceptual framework that allows for the proposal of hypotheses to explain the differentiation or specificity of venoms in an ecological and evolutionary context.

Understanding how these venoms have evolved in relation to interspecific interactions with humans would help to investigate the anthropogenic effect on natural populations of pitvipers, as well as to resolve some questions related to the effectiveness of antivenoms. Our goal in this section was to explain that snake venoms, before being a biological component that triggers a complex clinical picture (see Chapter 9), are a lethal hunting weapon modified and selected through the course of evolution of many species that grants the individual one of the most sophisticated natural resources for food and defense.

3.2. Studies in proteomics in Colombia

Despite the great diversity of pitviper snakes in Colombia, only nine species (40%) have been the object of proteomics studies of at least one population across their distribution range in the country (Table 2). The proteins present in these venoms are categorized into seven to twelve protein families. Generally, PLA₂s are the most abundant components identified, followed by SVMPs, SVSPs, and LAAOs. However, there are some exceptions. For instance, the venom of *C. durissus* exhibits a significant concentration of crotoxin (64.71%), which is a characteristic of South American rattlesnakes [170,176], but between Colombian populations there are intraspecific differences in which crotamine is only present in the Caribbean ecoregion [171].

Table 2. Percentage of toxins in the venom composition of the pitviper species distributed in Colombia with proteome characterization.

Toxins	<i>B. atrox</i>	<i>B. asper</i>	<i>B. ayerbel</i> **	<i>B. rhombeatus</i> **	<i>B. punctatus</i>	<i>P. lansbergii</i>	<i>B. myersi</i>	<i>L. acrochorda</i>	<i>C. durissus</i>
SVMPs	48.5	33.17	53.7	39.7	41.4	35.5	21.50	23.2	3.3
PLA ₂ s	24.1	31.29	0.7	23.0	9.3	16.2	54.04	2.3	0.58
SVSPs	10.9	3.89	9.3	4.9	5.4	4.5	3.43	35.1	6.33
LAAO	4.7	3	3.3	2.1	3.1	3.6	1.10	9.6	3.16
Hya	*	*	*	*	*	*	0.01	*	*
Crotoxin	*	*	*	*	*	*	*	*	64.71
CRISP	2.6	1.45	1.1	0.7	1.2	1.4	*	0.9	1.27
Dis	1.7	3.27	2.3	4	3.8	12.9	0.62	*	13.7
PLB	*	*	*	*	*	0.7	0.37	*	*
PDE	*	*	0.7	0.2	*	0.3	0.08	*	*
CTL	7.1	8.54	10.1	3.1	16.7	6.7	0.56	6.9	1.18
VEGF	*	*	*	*	1.7	2.2	*	0.6	*
NGF	*	*	0.1	*	*	*	0.07	*	*
PNP	*	*	*	*	*	*	*	*	*
BPP	0.3	*	8.3	7	*	*	*	21.5	*
PEP	*	*	8.7	*	10.7	*	9.07	*	*
Nuc	*	*	*	*	*	0.4	0.02	*	*
SVMPI	*	*	*	10	*	*	*	*	*
Crotamine	*	*	*	*	*	*	*	*	5.77

Abbreviations for protein family names: PLA₂s: phospholipase A₂; SVMPs: metalloproteinase; LAAO: L-amino acid oxidase; CTL: C-type lectin/lectin-like; CRISP: cysteine-rich secretory protein; SVSPs: serine proteinase; Nuc: nucleotidase; PDE: phosphodiesterase; Hya: hyaluronidase; NGF: nerve growth factor; PLB: phospholipase B; PNP: peptides and/or non-protein compounds; BPP: bradykinin-potentiating peptide. * Toxins no detected in the proteome. ** Dubious species of the *Bothrops asper* species complex (see Chapter 1) [118,136,171,181,184,187,192,195].

Until now, the venoms of *Bothrops* species had higher proportions of SVMPs than PLA₂. This same pattern was observed in the venoms of *P. lansbergii* and *L. acrochorda*. In contrast, the venom of *Bothrocophias myersi* has a superior percentage of PLA₂s. In addition, the venom of this species is the most complex, with twelve families of proteins. Special characteristics of *L. acrochorda* venom are their elevated amounts of SVSPs and BBPs (35.1% and 25.5%), even more than PLA₂s, which is a unique pattern. While the venom of the *L. muta* (Amazon bushmaster) has the highest quantity of LAAOs with 9.6%.

Interestingly, the venom of *B. punctatus* exhibits a higher concentration of C-type lectins (16.7%); however, these toxins were identified in all venoms through proteomic studies, ranging from 0.56% to 10.1% in the remaining species with proteomes available. It should be noted that the venom of *P. lansbergii* possesses the highest concentrations of desintegrins, whereas these toxins were not detected in the venom of *L. acrochorda*. One noteworthy discovery was the presence of hyaluronidases, the spreading factor of venoms (as discussed in Chapter 5) that were only detected in the venom of *B. myersi*, but in a minimal amount (0.01%). This result may be attributed to the fact that in certain instances not all proteins were classified. On average eight groups of toxins were described in most of the research, where percentages were usually informed for the following toxins: SVMPs, PLA₂s, SVSPs, LAAO, CRISP, Dis, CTL and BPP. Crotoxin and crotoamine are exclusive components for the *Crotalus* genus.

The venom of *Crotalus durissus* requires special attention because it is the only one with neurotoxic activity and low molecular weight myotoxins among Colombian viperids. The venom of Colombian rattlesnakes is composed of a considerable proportion of crotoxin (64.71%), the toxin responsible for inducing diaphragm-flaccid paralysis. However, another exceptional toxin present in this venom is crotoamine, a small basic polypeptide with myotoxic and cell-penetrating activities [177,178]. Interestingly, the expression of crotoamine can be different in venom from different geographical regions, showing an intraspecific variation in the composition of the *C. durissus* venom [171,179].

3.3 The biological activities of Colombian pitvipers

All venom activities evaluated for species from Colombia are correlated with the signs and symptoms observed in snakebite cases reported (see Chapter 9). Venoms from *Bothrops* species (*B. atrox*, *B. asper*, *B. ayerbeii*, *B. rhombeatus* and *B. punctatus*; see Figure 6) have different lethal activity values (LD₅₀ µg/mice), such as 81.4 (80.2–83.6) [42], 63 (50–81 ranges for several ecoregions from Colombia) [180], 50.1 (37.5–58.3) [181], 54.9 (36.0–83.8) [181], and 47 (36–61) [180]. Hence, the most lethal venoms among the *Bothrops* species tested so far are *B. asper* and *B. punctatus*. It is important to mention that not all Colombian *Bothrops* species have been evaluated. However, the venom of *C. durissus* is the most lethal venom within pitvipers in Colombia, with a lethal dose average of 1 µg/mice (0.02–2.5) [171,180]. This lethal activity value is considered a consequence

of the venom's neurotoxicity, a widely described functional feature that is biologically absent in the venoms of other pitviper species [182,183].

Saldarriaga et al. [42] carried out one of the pioneering studies on the ontogeny of *Bothrops* venoms that allowed us to understand how the species in its first stages of life develops venoms with fractions endowed with high hemorrhagic activity, while in the adult stage mostly fractions are found with indirect hemolytic activity. Furthermore, when *B. asper* venoms from Colombia and Costa Rica are compared, there are specific divergences associated with variations in lethal, hemorrhagic, edematogenic, myotoxic and indirect hemolytic activities [42].

In this study, we compared the biological activities of the principal biological important pitviper species from Colombia. We found that within the *B. asper* lineages (*B. asper*, *B. ayerbeii*, and *B. rhombeatus* [184]) the venom of main lineage of *B. asper* (distributed throughout the Caribbean region and the middle Magdalena Valley) is significantly more lethal than *B. ayerbeii*, and *B. rhombeatus* lineages. *Bothrops asper* venom turned out to be more hemorrhagic than venom of the *B. rhombeatus* lineage, but the minimum hemorrhagic dose of *B. ayerbeii* lineage was significantly lower, showing its greater capability to cause hemorrhage.

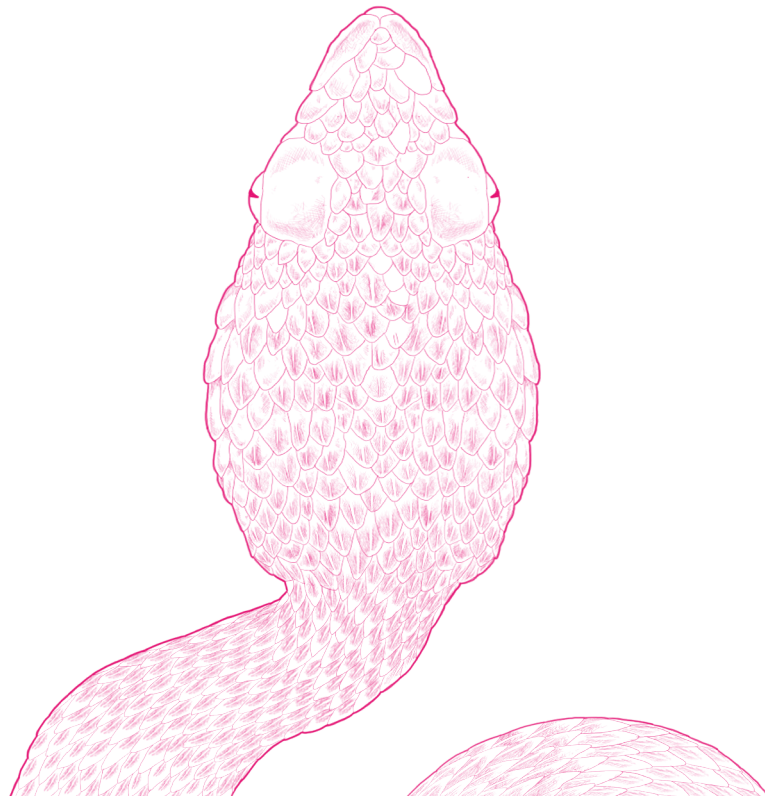
On the other hand, the venom of *B. rhombeatus* lineage showed a greater coagulating capacity while *B. ayerbeii* lineage exhibited the lowest coagulating capacity. Moreover, despite the geographic isolation between the populations of *B. asper* from Gorgona Island (Pacific Ocean) and the continental populations from the Valle del Cauca, the biological activities of their venoms showed very similar activities, with the exception of coagulating activity, which is much lower in the populations from Gorgona Island. However, the biological activities of the venom of these two populations are significantly different when compared to the venoms from the Caribbean coast and Magdalena River Basin [185]. It has been inferred that the divergence of populations within some lineages of *B. asper* is strongly influenced by the orography of the Andes mountain range, and the dynamically changing nature of their ecosystems that could drive specificity in different ecological niches [184]. In fact, a proteomic comparison of *B. asper* venoms based on the genetic diversity documented for lineages of the *Bothrops asper-atrox* complex found intraspecific variation in four of the main components of the venom [53,60]: SVMP, SVSP, PLA₂ and C-type lectin-like proteins (CTL) [184].

There is controversy regarding the biological activities of the venom of various populations of the *Bothriechis schlegelii* species complex. As we explained earlier, this species exhibits clinal variation among its populations, which is possibly related to the variability in the biological activities of their venoms (Figure 6A, C-E). For example, the Colombian populations from the central-eastern Andes (Antioquia department) [180,186], and those from the southwestern region of Colombia (Valle del Cauca department) [187] do not exhibit hemorrhagic activity. However, the venom of these populations shows coagulating activity and a high capacity for ede-

ma formation, similar to what is observed in the populations from Costa Rica (*Bothriechis nigroadspersus*) [188]. Given the medical importance of this species and its wide distribution along the agricultural production zones of the Colombian Andes, it is a priority to conduct rigorous studies exploring the determinants of the variability in the biological activities of the *Bothriechis schlegelii* species complex. This will enable physicians treating envenomations to generate more accurate diagnoses, establish timely and appropriate therapy, and anticipate possible clinical complications.

Hemorrhagic activity of snake venoms is attributed to the activity of SVMPs [189]. All pitviper venoms tested up till now have hemorrhagic activity; however, their potency is variable. Generally, *Bothrops* spp., *Porthidium* spp., *L. acrochorda* and *Bothrocophias* spp. are more hemorrhagic than the *B. schlegelii* species complex, whereas *C. durissus* rarely induces local hemorrhage [190–193]. Nevertheless, *Bothrocophias campbelli* venom induces mild hemorrhage and coagulation disorders [194].

Both *Porthidium lansbergii* and *P. nasutum* show medium cytotoxic activity and low lethality when assessed on mice, but both species show high hemorrhagic activity [195]. In addition, pitviper venoms from Colombian populations induce hemostatic disorders that are related to their pro-coagulant activity shown in vitro, their fibrinogenolytic displayed in vivo, and the alteration of coagulation times. These venoms are characterized by a massive consumption of fibrinogen that can only be recovered by antivenom therapy [90,193,196].



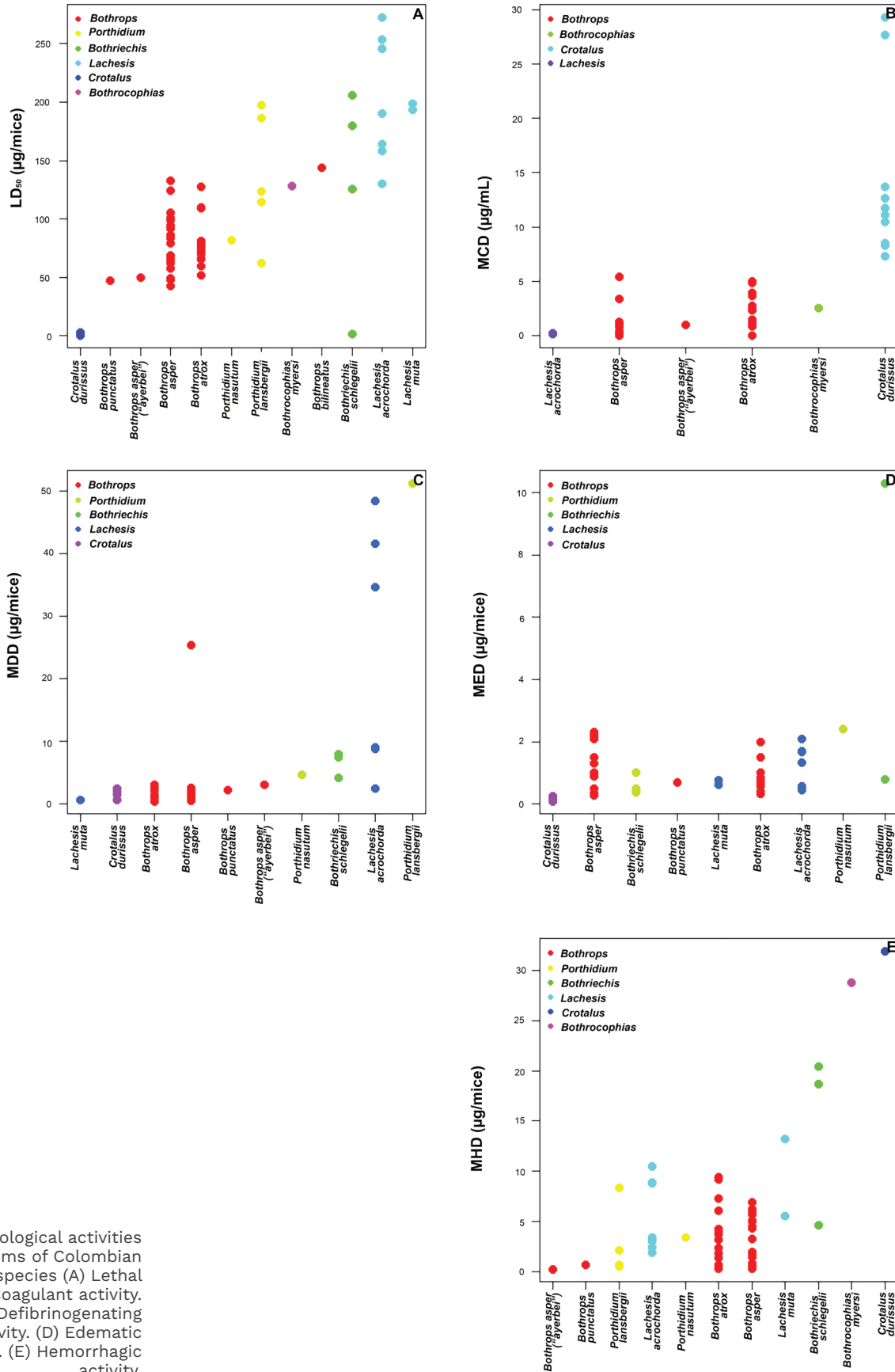


Figure 6. Biological activities of venoms of Colombian pitviper species (A) Lethal dose. (B) Coagulant activity. (C) Defibrinogenating activity. (D) Edematic activity. (E) Hemorrhagic activity.

Table 3. Biological activities tested for Colombian Viperidae

Species	Lethality	Hemorrhagic	Myotoxicity	Edematogenic	Coagulant	Hemolytic (PLA ₂ activity)	Defibrinogenating	Neurotoxic
<i>Bothriechis schlegelii</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Bothrocophias myersi</i>	Evaluated	Present	Present	Present	Present	Unevaluated	Unevaluated	Absent
<i>Bothrops asper</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
Lineage <i>B. ayerbei</i> *	Evaluated	Present	Present	Present	Present	Present	Present	Absent
Lineage <i>B. rhombeatus</i> *	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Bothrops atrox</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Bothrops punctatus</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Crotalus durissus</i>	Evaluated	Present	Present	Present	Present	Present	Present	Present
<i>Lachesis acrochorda</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Lachesis muta</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Porthidium lansbergii</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Porthidium nasutum</i>	Evaluated	Present	Present	Present	Present	Present	Present	Absent
<i>Bothrocophias campbelli</i>								
<i>Bothrocophias colombianus</i>								
<i>Bothrocophias hyoprora</i>								
<i>Bothrocophias myrringae</i>								
<i>Bothrocophias tulitoi</i>								
<i>Bothrops bilineatus</i>								
<i>Bothrops oligobalius</i>								
<i>Bothrops pulcher</i>								
<i>Bothrops taeniatus</i>								
<i>Bothrops venezuelensis</i>								

* Dubious species of the *Bothrops asper* species complex (see Chapter 1). [42,118,136,171,180-190,194]

3.4 Outlook for antivenom research

Antivenom therapy, (based on serum therapy; see Chapter 6), was developed in the 19th century; and although it has had huge improvements throughout its existence, this essentially uses immunoglobulins as therapy to this day [197]. In fact, since their production, antivenoms have been used successfully for the treatment of snakebite envenomation in various regions of the world [108]. In context, the antivenom “works” from purified immunoglobulins, which act initially by activating the immune response to the most pathogenic venom proteins, and then, trying to reduce and stop the severity of the envenomation. However, in some cases this therapy can cause adverse reactions (e.g., anaphylactic shock) [111].

The administration of antivenoms is done intravenously. Once in the bloodstream, the antibodies or antibody fragments bind to the venom toxins, preventing them from damaging tissues or interfere with critical bodily functions. These immunoglobulins are obtained and purified from the plasma of large mammals (usually horses), which have been previously immunized with sub-lethal doses of snake venom [198,199] (see Chapter 6). Antivenoms can be either monovalent or polyvalent, depending on whether the horses are immunized with venom from a single species or with a mixture of venoms from multiple species, respectively [200].

The incidence and severity of adverse reactions associated with the administration of antivenom have been reduced by the purification of immunoglobulins [201], thus demonstrating the therapeutic efficacy of antivenom to control the systemic manifestations of snake envenoming [202]. However, the venom heterogeneity between species could explain the differences in the clinical symptoms recorded from different geographic regions during snakebite cases [154]. This phenomenon reflects a series of local adaptations that confer a series of biological advantages on a certain population of snakes. Currently, snake envenoming represents a huge challenge for the design of new approaches to immunotherapy [111].

The description of venoms from various species and the knowledge of implicit inter- and intraspecific variability raises a scenario of new and necessary research approaches for the selection of phylogeographic regions of interest to help capture and describe venom variability. For example, neutralization measurements of the *C. durissus* venom using National Institute of Health of Colombia (INS) antivenom to calculate the median effective dose (ED_{50}) have shown that more antivenom is required to neutralize venom from Caribbean populations (2.3 mg venom/mL) than those from middle Magdalena basin (1.4 mg venom/mL), or the Orinoquia ecoregion (0.6 mg venom/mL) [171]. Therefore, there is a need to use venoms combined as a substrate to produce antivenom and, thus, prioritize the investigation of the snake venom using a combination of proteomic tools, functional toxicological and biochemical [199]. In fact, Juan Calvete et al. [111] presents a useful protocol to investigate antigen-antibody immunoreactivity, opening the door to the possible therapeutic utility of antivenoms against homologous and heterologous venoms in pitviper species.

Thus, based on the immuno-reactivity of the various toxins against antivenoms, these toxic components could be grouped into three types of toxins: completely immunodepletable, partially immunodepleted, and non-immunodepleted. Thus, immunization protocols must be improved by using immunogen mixtures [111]. This approach has recently established the basis for the development of antivenoms on an immunologically sound foundation and has proposed the use of immunochemical analysis based on proteomics for the identification of the most medically relevant venoms from snakes present in a specific region [111].

A first example of this type of new therapy, which has impacted and driven research in the field of antivenoms, is the development of an antivenom designed for the treatment of snakebites caused by *Bothrops lanceolatus* (endemic to the island of Martinique in the Lesser Antilles). This Sanofi-Pasteur Bothrofav®, has demonstrated good preclinical efficacy in neutralization, timely preventing the development of the most severe systemic events [203,204].

The limited availability of antivenoms in several Central and South American countries, as a result of various anthropogenic processes (social and environmental; see Chapter 6), has created the need and challenge to improve the neutralizing capacity of the available antivenoms. As an alternative, the use of small molecular inhibitors has been proposed to counteract the effect of the main snake venom proteins that cause clinical symptoms. Research in this field has recently gained relevance [144]. The effect of these inhibitors has been studied both in vitro and in vivo, yielding promising results for the inhibition of PLA₂, SVMP, and SVSP [205,206].

Researchers from the Clodomiro Picado Institute, led by Dr. José María Gutiérrez, conducted detailed proteomic studies of the venoms of *Bothrops caribbaeus* and *B. lanceolatus* (species included within the *B. atrox* complex), finding immunoreactivity to the polyvalent antivenom produced in Costa Rica to neutralize the venom of Central American rattlesnakes. This antivenom was able to immunodegrade 80% of the proteins and neutralize the lethal activities of the venoms in these two species [205]. Other studies highlight the importance of comparing proteomes between species with a similar natural history but with taxonomic uncertainties or undefined lineages within the *Bothrops asper-atrox* complex [111,207,208]. The findings of the Costa Rican researchers represent a significant advancement in understanding the cross-reactivity of antivenoms that counteract the symptoms of envenomation caused by American pit vipers.

In Colombia, experiments have been conducted to explore complementary options to antivenom treatment. Recently, recombinant proteins have been produced to improve the immunogenicity of proteins such as PLA₂ and three-finger toxins, representing a novel advancement for the future development of antivenoms that neutralize the toxins of snakes from the *Micrurus* genus [209]. Additionally, experiments using purified molecules from plant extracts to inhibit certain biological activities of snake venoms [192,210,211], as well as synthetic inhibitors that

have shown inhibitory activity against PLA₂ and SVMP [212] have all been carried out in preclinical trials.

The gradual incorporation of chromatographic and mass spectrometry techniques to complement the current platforms recommended by the WHO in the evaluation of antivenoms are ideal for establishing a geographical range of their clinical validity [213]. Besides, when combined with *in vivo* neutralization assays, antivenoms represent a good complement for evaluating the therapeutic efficacy of antivenoms against the same venoms used in its production [214].

Polyvalent antivenoms such as those from ICP (Costa Rica), INS (Colombia), PROBIOL (Colombia), UCV (Venezuela), BIOL (Argentina), and INS-PERU (Peru) can recognize the main toxin families present in *Bothrops asper* venoms (SVMP, PLA₂s, CRISP, SVSP, CTL); among these, the INS antivenom is the most effective and safest therapy [181]. In comparative studies of the antivenoms available in Colombia, the INS antivenom showed the most effective immunorecognition capacity, as well as the best neutralizing capacity against the biological activities of venoms from *Bothrops* spp., *Bothrocophias* spp., *Bothriechis* spp., *Crotalus durissus*, *Lachesis* spp., and *Porthidium* spp., followed by the Silanes (Bioclon) antivenom from Mexico [190]. In a more recent study [185], *B. asper* venoms from Gorgona Island and the Pacific and Western ecoregions of Colombia were evaluated from immunoreactivity assays using INS and Probiol antivenoms. The results obtained indicate again that the INS antivenom had higher antibody titers and neutralization compared to Probiol antivenom, showing the greater neutralizing capacity for the venoms of *B. asper* from Gorgona island population.

It is important to study the variability in the protein composition of different pitviper venoms and to develop studies that describe the structure-function correlations of individual toxins that will help to improve an understanding of venoms as trophic and defensive adaptations in different species [213]. Likewise, it is important to continue exploring alternatives for the potential use of venom toxins as active compounds in the development of new medications. However, this should be preceded by a deeper study of the natural history of viperid genera present in Colombia (e.g., distribution), to better understand how the destruction of their natural habitats and climate change affect the variability of their venoms and their survival in both natural and altered environments.

Appendix: Material and Methods

Phylogenetic analyses

Phylogenetic analyses were performed using maximum-likelihood estimation in the IQ tree software [215]. The evolutionary models were performed using the model finder [216]: TN+F+I: CYTB1_ND41, TIM+F+R2: CYTB2, GTR+F+I+G4: CYTB3_ND43, TN+F+R2: ND42. The analyses were performed using an array of 102 concatenated sequences for 51 terminals for the *Bothrops atrox* complex and the *Porthidium* genus from Colombia, Brazil, Ecuador, Venezuela, and Costa Rica. We used 20 species as out groups: *Gloydus halys*, *Sistrurus miliarius* and *S. catenatus*; *Crotalus horridus*, *C. ruber*, *C. atrox*, and *C. durissus*; *Agkistrodon piscivorus* and *A. contortrix*; *Mixcoatlus melanurus*; *Ophryacus ondulatus*; *Cerrophidion godmani*; *Metlapilcoatlus (Atropoides) nummifer* and *M. mexicanus*; *Atropoides picadoi* and *Bothrocophias hyoprora*, *B. tulitoi* and *B. myrringae*; *Bothriechis schlegelii* (currently redescribed as *Bothriechis nigroadspersus*) and *Bothrops jararaca*. Besides, phytools were used to perform a relationship analysis (heatmap) at the phylogenetic level [217], using two published proteomes for Colombian pitviper species and some related lineages/clades.

Biological activity assays

The biological assays for lethality, coagulant, defibrinogenating, hemorrhagic, and edematogenic activity were conducted by the INS (National Institute of Health) as part of its antivenom production activities, in accordance with venom characterization guidelines established by the WHO in "Guidelines for the Production, Control and Regulation of Snake Antivenom Immunoglobulins; 2017". All tests were performed on CD1 strain mice weighing 18-22 g, following INS protocols for the use of laboratory animals.

Median Lethal Dose (LD₅₀).— We assessed venom lethality following the procedures and protocols described by [200,218], as well as the INS production protocol MEN-R04.6022-011. The median lethal dose (LD₅₀) of a venom is defined as the amount of venom that causes death in 50% of inoculated mice. To determine the LD₅₀, different doses of venom were prepared in saline solution and administered intraperitoneally to groups of five mice. Mortality was recorded after 48 hours. A control group of five mice received saline solution. Finally, the LD₅₀ was estimated using statistical methods such as the Spearman-Kärber, Probit function, or other non-parametric methods [219].

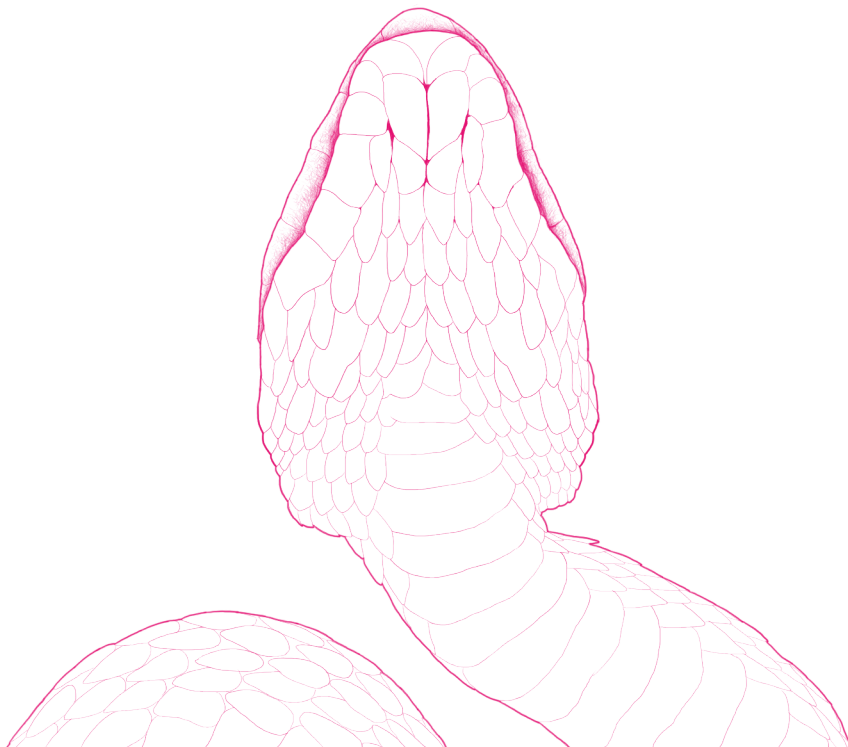
Minimum Coagulant Dose (MCD).— In vitro coagulation assays were performed following the procedures and protocols described by [200,220], as well as the INS production protocol MEN-R04.6031-001. The minimum coagulant dose (MCD) was defined as the lowest venom dose that induced coagulation in citrated human plasma.

Minimum Defibrinogenating Dose (MDD).— In vivo defibrinogenation assays were conducted following the INS production protocol MEN-R04.6022-046, as well as the procedures and protocols described by [200,220]. The MDD corresponds to the venom dose that, when adminis-

tered intravenously to five mice injected with a venom dilution, induces incoagulability after 2 hours of sample collection.

Minimum Edematogenic Dose (MED).— In vivo edema assays were performed following the INS production protocol MEN-R04.6022-043, as well as the procedures and protocols described by [200]. The MED is defined as the venom dose that induces a 30% increase in mouse paw edema following the injection of a venom dilution. For this purpose, venom solutions at different concentrations were prepared. Each dilution was injected into the right hind paw plantar pad of five mice, while the left paw received sterile saline (0.85%). One hour after inoculation, the animals were ethically euthanized [221]. Both paws (at the tibiotarsal joint) were excised and weighed.

Minimum Hemorrhagic Dose (MHD).— In vivo hemorrhagic assays were conducted following the INS production protocol MEN-R04.6022-049, as well as the procedures and protocols described by [200,220]. The minimum hemorrhagic dose (MHD) was defined as the smallest amount of venom capable of inducing a hemorrhagic lesion with a 10 mm diameter. To determine this, different venom dilutions in saline solution were prepared, with a final injection volume of 50 μ L. Each dilution was then injected intradermally into the shaved ventral skin of five mice. After 2-3 hours, the animals were ethically euthanized [221]. The skin was dissected, and the diameter of the resulting hemorrhagic lesion was measured.



References

1. Cubides-Cubillos, S.D. Actitudes, Creencias y Prácticas Desarrolladas En Comunidades Rurales Del Urabá Antioqueño Con Relación a Las Serpientes y Su Mordedura. Trabajo presentado para optar al título de Magister en Educación – Línea de formación: pedagogía y diversidad cultural, Universidad de Antioquia: Medellín, Colombia, **2016**.
2. Campbell, J.A.; Lamar, W. *The Venomous Reptiles of the Western Hemisphere*; Cornell University Press, Ed; Comstock Publishing: Ithaca, New York, **2004**.
3. Alencar, L.R.V.; Quental, T.B.; Graziotin, F.G.; Alfaro, M.L.; Martins, M.; Venzon, M.; Zaher, H. Diversification in Vipers: Phylogenetic Relationships, Time of Divergence and Shifts in Speciation Rates. *Molecular Phylogenetics and Evolution* **2016**, *105*, 50–62, doi:10.1016/j.ympev.2016.07.029.
4. Uetz, P.; Freed, P.; Aguilar, R.; Hošek, J. The Reptile Database. **2023**.
5. Palisot de Beauvois Memoir on Amphibia. Serpents. *Transactions of the American Philosophical Society* **1799**, *4*, 362–381.
6. Werman, S.D. Phylogenetic Relationships of Central and South American Pitvipers of the Genus *Bothrops* (Sensu Lato): Cladistic Analyses of Biochemical and Anatomical Characters. In *Biology of the Pitvipers*; J.A. Campbell, E.D. Brodie, Eds.; Selva: Texas, **1992**; pp. 21–40.
7. Peters, W. Über Die von Hrn. Dr. Hoffmann in Costa Rica Gesammelten Und an Das Königl. Zoologische Museum gesandten Schlangen **1859**, pp. 275–278.
8. Gutberlet, R.L.; Campbell, J.A. Generic Recognition for a Neglected Lineage of South American Pitvipers (Squamata: Viperidae: Crotalinae), with the Description of a New Species from the Colombian Choco. *American Museum Novitates* **2001**, 1–15.
9. Wagler, J. Serpentina Brasiliensium Species Novae, Ou Histoire Naturelle Des Espèces Nouvelles de Serpens. *Jean de Spix, Animalia nova sive species novae* **1824**, *VII*, 1–75.
10. Campbell, J.A.; Lamar, W.W. Taxonomic Status of Miscellaneous Neotropical Viperids, with the Description of a New Genus. *Occasional Papers, Museum of Texas Tech University*. **1992**, *153*, 1–31.
11. Linnaeus, C. *Systema Naturæ per Regna Tria Naturæ, Secundum Classes, Ordines, Genera, Species. Cum Characteribus, Differentiis, Synonymis, Locis*; 10th ed.; **1758**; Vol. 1.
12. Daudin, F.M. Histoire Naturelle, Générale et Particulière, *Des Reptiles*; F. Dufart.; Paris, **1802**; Vol. 4.
13. Campbell, J.D.; Frost, R.; and Castoe, T.A. A new generic name for jumping pitvipers (Serpentes: Viperidae). *Revista Latinoamericana de Herpetología* **2019**, *2*, 52–53.
14. Jadin, R.C.; Gutberlet Jr, R.L.; Smith, E.N. Phylogeny, Evolutionary Morphology, and Hemipenis Descriptions of the Middle American Jumping Pitvipers (Serpentes: Crotalinae: *Atropoides*). *Journal of Zoological Systematics and Evolutionary Research* **2010**, *48*, 360–365, doi:10.1111/j.1439-0469.2009.00559.x.
15. Cope, E.D. Catalogue of Batrachians and Reptiles [Batrachia and Reptilia] of Central America and Mexico. **1887**, *32*, 1–98.

16. Cope, E.D. "Ninth Contribution to the Herpetology of Tropical America." *Proceedings of the Academy of Natural Sciences of Philadelphia* **1871**, 23, 200–224.
17. Garman, S. The Reptiles and Batrachians of North America. *Memoirs of the Museum of Comparative Zoölogy*, Cambridge (Massachusetts) **1884**, 8, 34–185.
18. Cope, E.D. «Notes upon Some REPTILES of the Old World». *Proceedings of the Academy of Natural Science of Philadelphia*. **1862**, 14, 337–344.
19. Boulenger, G.A. "An Account of the Reptilia Obtained in Burma, North of Tenasserim, by Monsieur. L. Fea, of the Genoa Civic Museum." *Annali del Museo Civico di Storia Naturale di Genova* **1888**, 6, 593–604.
20. Gray, J.E. Monographic Synopsis of the Vipers, or the Family Viperidæ. *Zoological Miscellany*, London **1842**, 2, 68–71.
21. Cope, E.D. Catalogue of the Venomous Serpents in the Museum of the Academy of Natural Sciences of Philadelphia, with Notes on the Families, Genera and Species. *Proceedings of the Academy of Natural Sciences of Philadelphia* **1860**, 11, 332–347.
22. Wagler, J.G. Natürliches System Der Amphibien, Mit Vorangehender Classification Der Säugetiere Und Vögel. *Ein Beitrag zur vergleichen Zologie* **1830**, 1–352.
23. Kuhl, K.; Van Hasselt, J.C. Uittreksel Uit de Brieven van Heeren Kuhl En van Hasselt, Aan de Heeren C.J. Temminck, Th. van Swinderen En W. de Haan. *Algemeene Konst. Letterbode* **1822**, 7, 99–104.
24. Gloyd, H.K. "A New Generic Name for the Hundred-Pace Viper." *Proceedings of the Biological Society of Washington* **1979**, 91, 963–964.
25. Merrem, B. Versuch Eines Systems Der Amphibien. *Tentamen Systematis Amphibiorum*. Marburg: J.C. Krieger **1820**, 191.
26. Alcock, A., F.F. "An Account of the Reptilia Collected by Dr. F. P. Maynard, Captain A. H. McMahon, C.I.E., and the Members of the Afghan-Baluch Boundary Commission of 1896." *Journal of the Asiatic Society of Bengal* **1897**, 65, 550–556.
27. Malhotra, A.; Thorpe, R.S. A Phylogeny of Four Mitochondrial Gene Regions Suggests a Revised Taxonomy for Asian Pitvipers (*Trimeresurus* and *Ovophis*). *Molecular Phylogenetics and Evolution* **2004**, 32, 83–100, doi:10.1016/j.ympev.2004.02.008.
28. Hoge, A.R.; Romano-Hoge, S.A.R. Poisonous Snakes of the World. Part 1: Check List of the Pitvipers, Viperioidea, Viperidae, Crotalinae. *Memorias do Instituto Butantan* **1981**, 42, 179–309.
29. Fitzinger, L. Systema Reptilium. Fasciculus Primus: Amblyglossae. *Vindobonae: Braumüller und Seidel* **1843**, 1–106.
30. Reuss, A.F. Sechs Europäische Giftschlangengattungen. *Zoologischer Anzeiger* **1927**, 75, 124–129.
31. Broadley, D.G. A Review of the Tribe Atherini (Serpentes: Viperidae), with Descriptions of Two New Genera. *African Journal of Herpetology* **1996**, 45, 40–48.
32. Nilson, G., T.B., A.C., O.N., J.U. & H.H.W. Taxonomic Position of the *Vipera xanthina* Complex. *Kaupia* **1999**, 8, 99–102.
33. McDiarmid, R.W., C.J.A., T.T. Snake Species of the World: A Taxonomic and Geographic Reference. *Herpetologists' League* **1999**, 1, 511.

34. Hoge A.R.; Romano-Hoge S.A.L. «Notes on Micro and Ultrastructure of ‘Oberhäutschen’ in Viperioidea. Viperioidea, Viperidae, Crotalinae». *Memorias do Instituto Butantan* **1983**, *44*, 81–118.
35. Boulenger, G.A. Catalogue of the Snakes in the British Museum (Natural History. Volume III., Containing the ... Viperidæ. *London: Trustees of the British Museum (Natural History). (Taylor and Francis, printers)* **1896**, *3*, 772.
36. Lacepede, B.G.E.L. Memoire Sur Plusieurs Animaux de La Nouvelle Hollande Dont La Description n’a Pas Encore Ete Publiee. *Annales du Museum d’Histoire naturelle* **1804**, *4*, 184–211.
37. Laurenti, J. Specimen Medicus Exhibens Synopsin Reptilium. *J. Thoma* (Vienna) **1768**, 1–2.
38. Wüster, W.; Peppin, L.; Pook, C.E.; Walker, D.E. A Nesting of Vipers: Phylogeny and Historical Biogeography of the Viperidae (Squamata: Serpentes). *Molecular Phylogenetics and Evolution* **2008**, *49*, 445–459, doi:10.1016/j.ympev.2008.08.019.
39. Greene, H. The Ecological and Behavioral Context for Pitviper Evolution. In *Biology of the Pitvipers*; **1992**; pp. 107–117.
40. Mackessy, S. Venom Ontogeny in the Pacific Rattlesnakes *Crotalus viridis helleri* and *C. d. oreganus*. *Copeia* **1988**, *2*, 92–101.
41. Guércio, R.A.P.; Shevchenko, A.; Shevchenko, A.; López-Lozano, J.L.; Paba, J.; Sousa, M. V.; Ricart, C.A.O. Ontogenetic Variations in the Venom Proteome of the Amazonian Snake *Bothrops atrox*. *Proteome Science* **2006**, *4*, doi:10.1186/1477-5956-4-11.
42. Saldarriaga, M.M.; Otero, R.; Núñez, V.; Toro, M.F.; Díaz, A.; Gutiérrez, J.M. Ontogenetic Variability of *Bothrops atrox* and *Bothrops asper* Snake Venoms from Colombia. *Toxicon* **2003**, *42*, 405–411, doi:10.1016/S0041-0101(03)00171-5.
43. Alape-Girón, A.; Sanz, L.; Escolano, J.; Flores-Díaz, M.; Madrigal, M.; Sasa, M.; Calvete, J.J. Snake Venomics of the Lancehead Pitviper *Bothrops asper*. Geographic, Individual, and Ontogenetic Variations. *Journal of Proteome Research* **2008**, *7*, 3556–3571, doi:10.1021/pr800332p.
44. Kardong, K.; Kiene, T.; Bels, V. Evolution of Trophic Systems in Squamates. *Neth Journal Zoology* **1997**, *47*, 1–17.
45. Burger, W.L. Genera of Pitvipers (Serpentes: Crotalidae), University of Kansas: Lawrence, **1971**.
46. Avise, J.C. *Molecular Markers, Natural History and Evolution*; Springer US, **1994**.
47. Avise, J.C. *Phylogeography*; Harvard University Press, **2000**; ISBN 9780674268708.
48. Zamudio, K.R.; Greene, H.W.; Zamudio, K.R.; Greene, H.W. Phylogeography of the Bushmaster (*Lachesis Muta*: Viperidae): Implications for Neotropical Biogeography. *Systematics and Conservation*; **1997**; Vol. 62.
49. Thorpe, R.S.; Mcgregor, D.P.; Cumming, A.M.; Jordan, W.C. DNA Evolution and Colonization Sequence of Island Lizards in Relation to Geological History: MtDNA RFLP, Cytochrome b, Cytochrome Oxidase, 12s RRNA Sequence, and RAPD Nuclear Analysis. *Evolution* (NY) **1994**, *48*, 230–240.
50. Parkinson, C.L. Molecular Systematics and Biogeographical History of Pitvipers as Determined by Mitochondrial Ribosomal DNA Sequences. *Copeia* **1999**, 576–586.

51. Gutberlet Jr, R.L.; Harvey, M.B. Phylogenetic Relationships of New World Pit Vipers as Inferred from Anatomical Evidence. In *Biology of the Vipers*; G.W. Schuett, M. Höggren, M.E. Douglas, H.W. Greene, Eds.; Eagle Mountain Publishing: Salta Lake City, **2002**; Vol. 1, pp. 51–68.
52. Amaral, A. Contribuicao ao conhecimento dos ofidios neotropicos XXXVI. Redescricao da espécie *Bothrops hyoprora* Amaral, **1935**. Mem. Inst. Butantan 26: 221-225.
53. Saldarriaga-Cordoba, M.; Parkinson, C.L.; Daza, J.M.; Wüster, W.; Sasa, M. Phylogeography of the Central American Lancehead *Bothrops asper* (Serpentes: Viperidae). *PLoS One* **2017**, *12*, e0187969, doi:10.1371/journal.pone.0187969.
54. Gregory-Wodzicki, K.M. Uplift History of the Central and Northern Andes: A Review. *Bulletin of the Geological Society of America* **2000**, *112*, 1091–1105, doi:10.1130/0016-7606(2000)112<1091:UHOTCA>2.3.CO;2.
55. Carrasco, P.A.; Mattoni, C.I.; Leynaud, G.C.; Scrocchi, G.J. Morphology, Phylogeny and Taxonomy of South American Bothropoid Pitvipers (Serpentes, Viperidae). *Zoologica Scripta* **2012**, *41*, 109–124, doi:10.1111/j.1463-6409.2011.00511.x.
56. Bagley, J.C.; Johnson, J.B. Phylogeography and Biogeography of the Lower Central American Neotropics: Diversification between Two Continents and between Two Seas. *Biological Reviews* **2014**, *89*, 767–790, doi:10.1111/brv.12076.
57. Carrasco, P.A.; Koch, C.; Grazziotin, F.G.; Venegas, P.J.; Chaparro, J.C.; Scrocchi, G.J.; Salazar-Valenzuela, D.; Leynaud, G.C.; Mattoni, C.I. Total-evidence Phylogeny and Evolutionary Morphology of New World Pitvipers (Serpentes: Viperidae: Crotalinae). *Cladistics* **2023**, *39*, 71–100, doi:10.1111/cla.12522.
58. Antonelli, A.; Quijada, A.; Crawford, A.J.; Bates, J.M. Molecular Studies and Phylogeography of Amazonian Tetrapods and Their Relation to Geological and Climatic Models Origin, Evolution, Genetics and Selective Drivers of Snake Venom Evolution. *View Project Population Genetics and Molecular Ecology of Ectothermic Vertebrates* **2010**.
59. Lagomarsino, L.P.; Condamine, F.L.; Antonelli, A.; Mulch, A.; Davis, C.C. The Abiotic and Biotic Drivers of Rapid Diversification in Andean Bellflowers (Campanulaceae). *New Phytologist* **2016**, *210*, 1430–1442, doi:10.1111/nph.13920.
60. Salazar-Valenzuela, D.; Kuch, U.; Torres-Carvajal, O.; Valencia, J.H.; Gibbs, H.L. Divergence of Tropical Pitvipers Promoted by Independent Colonization Events of Dry Montane Andean Habitats. *Journal of Biogeography* **2019**, *46*, 1826–1840, doi:10.1111/jbi.13661.
61. Solís-Lemus, C.; Ané, C. Inferring Phylogenetic Networks with Maximum Pseudolikelihood under Incomplete Lineage Sorting. *PLoS Genetics* **2016**, *12*, doi:10.1371/journal.pgen.1005896.
62. Sousa, V.; Hey, J. Understanding the Origin of Species with Genome-Scale Data: Modelling Gene Flow. *Nature Reviews Genetics* **2013**, *14*, 404–414, doi:10.1038/nrg3446.
63. Wüster, W.; Salomão, M.; Quijada-Mascareñas, J.A.; Thorpe, R.; BBBSP Origins and Evolution of the South American Pitviper Fauna: Evidence from Mitochondrial DNA Sequence Analysis. In Proceedings

- of the Biology of the Vipers; Campbell, J.A., Brodie, E.D., Schuett, G.W., Hoggren, M., Douglas, M.E., Greene, H.W., Eds.; Eagle Mountain Publishing: UT, USA, **2002**; pp. 111–128.
64. Díaz-Ricaurte, J.; Cubides-Cubillos, S.D.; Ferreto Fiorillo, B. *Bothrops asper* (Garman, 1884). *Catálogo de Anfibios y Reptiles de Colombia* **2018**, *4*, 4–22.
 65. Cubides-Cubillos, S.D.; Loaiza-López, F.; Molina-Betancourt, J. *Porthidium Nasutum* (Bocourt, 1868). *Catálogo de Anfibios y Reptiles de Colombia* **2021**, *7*, 64–73.
 66. Molina-Betancourth, J.; Loaiza-López, F.; Cubides-Cubillos, S.D. *Porthidium Lansbergii* (Schlegel, 1841). *Catálogo de Anfibios y Reptiles de Colombia* **2021**, *7*, 51–63.
 67. Gutenkunst, R.N.; Hernandez, R.D.; Williamson, S.H.; Bustamante, C.D. Inferring the Joint Demographic History of Multiple Populations from Multidimensional SNP Frequency Data. *PLoS Genetics* **2009**, *5*, doi:10.1371/journal.pgen.1000695.
 68. Pickrell, J.K.; Pritchard, J.K. Inference of Population Splits and Mixtures from Genome-Wide Allele Frequency Data. *PLoS Genetics* **2012**, *8*, doi:10.1371/journal.pgen.1002967.
 69. Rittmeyer, E.N.; Austin, C.C. Combined Next-Generation Sequencing and Morphology Reveal Fine-Scale Speciation in Crocodile Skinks (Squamata: Scincidae: Tribolonotus). *Molecular Ecology* **2015**, *24*, 466–483, doi:10.1111/mec.13030.
 70. Leaché, A.D.; Banbury, B.L.; Felsenstein, J.; De Oca, A.N.M.; Stamatakis, A. Short Tree, Long Tree, Right Tree, Wrong Tree: New Acquisition Bias Corrections for Inferring SNP Phylogenies. *Systematic Biology* **2015**, *64*, 1032–1047, doi:10.1093/sysbio/syv053.
 71. McCormack, B.; Rycroft-Malone, J.; DeCorby, K.; Hutchinson, A.M.; Bucknall, T.; Kent, B.; Schultz, A.; Snelgrove-Clarke, E.; Stetler, C.; Titler, M.; et al. A Realist Review of Interventions and Strategies to Promote Evidence-Informed Healthcare: A Focus on Change Agency. *Implementation Science* **2013**, *8*, doi:10.1186/1748-5908-8-107.
 72. Pyron, R.A.; Costa, G.C.; Patten, M.A.; Burbrink, F.T. Phylogenetic Niche Conservatism and the Evolutionary Basis of Ecological Speciation. *Biological Reviews* **2015**, *90*, 1248–1262, doi:10.1111/brv.12154.
 73. Shanker, K.; Vijayakumar, S.P.; Ganeshaiyah, K.N. Unpacking the Species Conundrum: Philosophy, Practice and a Way Forward. *Journal of Genetics* **2017**, *96*, 413–430, doi:10.1007/s12041-017-0800-0.
 74. Mallik, A.K.; Srikanthan, A.N.; Ganesh, S.R.; Vijayakumar, S.P.; Campbell, P.D.; Malhotra, A.; Shanker, K. Resolving Pitfalls in Pit Viper Systematics - A Multi-Criteria Approach to Species Delimitation in Pitvipers (Reptilia, Viperidae, Craspedocephalus) of Peninsular India Reveals Cryptic Diversity. *Vertebrate Zoology* **2021**, *71*, 577–619, doi:10.3897/vz.71.e66239.
 75. Angarita-Sierra, T.; Cubides-Cubillos, S.D.; Hurtado-Gómez, J.P. Hidden in the Highs: Two New Species of the Enigmatic Toadheaded Pitvipers of the Genus *Bothrocophias*. *Vertebrate Zoology* **2022**, *72*, 971–996, doi:10.3897/vz.72.e87313.
 76. Haffer, J. Speciation in Amazonian Forest Birds. *Science* **1969**, *165*, 131–137.
 77. Fjelds, J. Geographical Patterns for Relict and Young Species of Birds

- in Africa and South America and Implications for Conservation Priorities; **1994**; Vol. 3.
78. Duellman, W.E. Global Distribution of Amphibians: Patterns, Conservation and Future Challenges. In *Patterns of distribution of amphibians: A global perspective*; Duellman, W.E., Ed.; John Hopkins University Press, **1999**; pp. 1–30.
 79. Myers, N.; Mittermeier, R.A.; Mittermeier, C.G.; Da Fonseca, G.A.B.; Kent, J. *Biodiversity Hotspots for Conservation Priorities*; **2000**; Vol. 403.
 80. Brumfield, R.T.; Edwards, S. V. Evolution into and out of the Andes: a bayesian analysis of historical diversification in *Thamnophilus antshrikes*. *Evolution* (NY) **2007**, *61*, 346–367, doi:10.1111/j.1558-5646.2007.00039.x.
 81. Coates, A.; Obando, J. “The Geologic Evolution of the Central American Isthmus.” In *Evolution and Environment in Tropical America*; Jackson, J., Budd, A., Coates, A., Eds.; The University of Chicago Press: Chicago, **1996**; pp. 21–56.
 82. Harvey, M.G.; Brumfield, R.T. Genomic Variation in a Widespread Neotropical Bird (*Xenops minutus*) Reveals Divergence, Population Expansion, and Gene Flow. *Molecular Phylogenetics and Evolution* **2015**, *83*, 305–316.
 83. Burbrink, F.T.; Castoe, T. Molecular Phylogeography of Snakes Rattlesnake Phylogeonomics and Morphology in MX: *Crotalus atrox* and *Crotalus scutulatus*. *View Project*; **2009**.
 84. Mason-Gamer, R.J.; Kellogg, E.A. Testing for phylogenetic conflict among molecular data sets in the tribe Triticeae (Gramineae); **1996**; Vol. 45.
 85. van Veller, M.G.P.; Zandee, M.; Kornet, D.J. Two Requirements for Obtaining Valid Common Patterns under Assumptions Zero, 1 and 2 in Vicariance Biogeography. *Cladistics* **1999**, *15*, 393–406.
 86. Crisci, J. V.; Katinas, L.; Posadas, P. Historical Bioeogeography: An Introduction; Harvard University Press: Cambridge, **2003**.
 87. Guarnizo, C.E.; Amézquita, A.; Bermingham, E. The Relative Roles of Vicariance versus Elevational Gradients in the Genetic Differentiation of the High Andean Tree Frog, *Dendropsophus labialis*. *Molecular Phylogenetics and Evolution* **2009**, *50*, 84–92, doi:10.1016/j.ympev.2008.10.005.
 88. Martins, M.; Marques, O.; Sazima, I. Ecological and Phylogenetic Correlates of Feeding Habits in Neotropical Pitvipers of the Genus *Bothrops*. In *Biology of the Vipers*; G.W. Schuett, M. Hoggren, M.E. Douglas, H.W. Greene, Eds.; Eagle Mountain Publishing, **2002**; pp. 307–328.
 89. Lynch, J.D. El Contexto de Las Serpientes de Colombia Con Un Análisis de Las Amenazas En Contra de Su Conservación. *Revista de la academia colombiana de Ciencias Exactas, Físicas y Naturales* **2012**, *36*, 435–449.
 90. Cañas-Dávila, C.A.; Castro-Herrera, F.; Castaño-Valencia, R.S. Serpientes Venenosas: Lecciones Aprendidas Desde Colombia; Tobón García, G.J., Osorno Reyes, J., Cañas Dávila, C.A., Castro Herrera, F., Castaño Valencia, R.S., Eugenia Rebolled, V., Eds.; Fundación Valle del Lili: Cali, Colombia., **2016**; Vol. 1.

91. Quiñones-Betancourt, E.; Díaz-Ricaurte, Juan.; Angarita-Sierra, T.; Guevara- Molina, C.; Díaz-Morales, R. *Bothrops Atrox* (Linnaeus, 1758). *Catálogo de Anfibios y Reptiles de Colombia* **2018**, 4, 7–23.
92. Salomão, M.G.; Wüster, W.; Thorpe, R.S. MtDNA Phylogeny of Neotropical Pitvipers of the Genus *Bothrops* (Squamata: Serpentes: Viperidae). *Kaupia (Darmstadt)* **1999**, 8, 127–134.
93. Wuster, W.; Salomao, M.G.; Duckett, D.; Thorpe, R.S. Systematics of the *Bothrops atrox* Complex: New Insights from Multivariate Analysis and Mitochondrial DNA Sequence Information. *Symposia of the Zoological Society of London* **1997**, 70, 99–113.
94. Castoe, T.A.; Parkinson, C.L. Bayesian Mixed Models and the Phylogeny of Pitvipers (Viperidae: Serpentes). *Molecular Phylogenetics and Evolution* **2006**, 39, 91–110, doi:10.1016/j.ympev.2005.12.014.
95. Castoe, T.A.; Sasa, M.M.; Parkinson, C.L. Modeling Nucleotide Evolution at the Mesoscale: The Phylogeny of the Neotropical Pitvipers of the Porthidium Group (Viperidae: Crotalinae). *Molecular Phylogenetics and Evolution* **2005**, 37, 881–898, doi:10.1016/j.ympev.2005.05.013.
96. Parkinson, C.L.; Zamudio, K.R.; Greene, H.W. Phylogeography of the Pitviper Clade *Agkistrodon*: Historical Ecology, Species Status, and Conservation of Cantils. *Molecular Ecology* **2000**, 9, 411–420.
97. Gibbs, H.L.; Sovic, M.; Amazonas, D.; Chalkidis, H.; Salazar-Valenzuela, D.; Moura-Da-Silva, A.M. Recent Lineage Diversification in a Venomous Snake through Dispersal across the Amazon River. *Biological Journal of the Linnean Society* **2018**, 123, 651–665, doi:10.1093/biolinnean/blx158.
98. Sandner-Montilla, F. Una Nueva Especie de Genero *Bothrops* (Serpentes, Crotalidae, Bothropinae) de La Region de Guanare, Estado Portuguesa, Venezuela. *Memorias científica de ofidiologia* **1979**, 4, 1–19.
99. Hallowell, E. Descriptions of Reptiles from South America, Supposed to Be New. *Proceedings of the Academy of Natural Sciences of Philadelphia* **1845**, 2, 241–247.
100. Markezich, A.L.; Taphorn, D.C. A Variational Analysis of Populations of *Bothrops* (Serpentes: Viperidae) from Western Venezuela. *Journal of Herpetology* **1993**, 27, 248–254.
101. Fenwick, A.M.; Gutberlet Jr, R.L.; Evans, J.A.; Parkinson, C.L. Morphological and Molecular Evidence for Phylogeny and Classification of South American Pitvipers, Genera *Bothrops*, *Bothriopsis*, and *Bothrocophias* (Serpentes: Viperidae). *Zoological Journal of the Linnean Society* **2009**; 156(3):617 – 640. DOI:10.1111/j.1096-3642.2008.00495.x
102. de Queiroz, K. “The General Lineage Concept of Species, Species Criteria, and the Process of Speciation.” In book: *Endless Forms: Species and Speciation* **1998**, 57–75.
103. Nascimento, D. dos S. Filogenia Molecular de Serpentes Neotropicais Do Grupo *Bothrops atrox* (Linnaeus, 1758) (Viperidae: Crotalinae). Tese de doutorado; **2014**.
104. Parkinson, C.L.; Campbell, J.A.; Chippindale, P. Multigene Phylogenetic Analyses of Pitviper Relationships, with Comments on Their Biogeography. In *Biology of the Vipers*; G.W. Schuett, M. Höggren, H.W. Greene, Eds.; *Biological Sciences Press*, Traverse City, **2002**; pp. 3–110.

105. Folleco-Fernández, A.J. Taxonomía del complejo *Bothrops asper* (serpentes: viperidæ) en el sudoeste de Colombia. Revalidación de la especie *Bothrops rhombeatus* (García 1896) y descripción de una nueva especie. *Revista Novedades Colombianas* **2010**, *10*, 33–70.
106. Ramírez-Chaves, H.E.; Solari, S. *Bothrops ayerbei* Folleco-Fernández, 2010 y *Bothrops rhomboatus* García, 1896 (Serpentes: Viperidae) Son Un Nombre No Disponible y Un Nomen Dubium, Respectivamente. *Boletín Científico Centro de Museos Museo de Historia Natural, Universidad del Caldas* **2014**, *18*, 138–141.
107. Garcia, E. Los Ofidios Venenosos Del Cauca. Métodos Empíricos y Racionales Empleados Contra Los Accidentes Producidos Por La Mordedura de Esos Reptiles. *Calí: Librería Colombiana* **1896**, 102.
108. Gutiérrez, J.M.; Calvete, J.J.; Habib, A.G.; Harrison, R.A.; Williams, D.J.; Warrell, D.A. Snakebite Envenoming. *Nature Reviews Disease Primers* **2017**, *3*, doi:10.1038/NRDP.2017.63.
109. Gutiérrez, J.M. Comprendiendo Los Venenos de Serpientes: 50 Años de Investigaciones En América Latina; *Revista de Biología Tropical* **2002**; Vol. 50.
110. Daltry, J.C.; Wüster, W.; Thorpe, R.S. Diet and Snake Venom Evolution. *Nature* **1996**, *379*, doi:10.1038/379537a0.
111. Calvete, J.J.; Sanz, L.; Angulo, Y.; Lomonte, B.; Gutiérrez, J.M. Venoms, Venomics, Antivenomics. *FEBS Letters* **2009**, *583*, 1736–1743, doi:10.1016/j.febslet.2009.03.029.
112. Gutiérrez, J.M.; Sanz, L.; Escolano, J.; Fernández, J.; Lomonte, B.; Angulo, Y.; Rucavado, A.; Warrell, D.A.; Calvete, J.J. Snake Venomics of the Lesser Antillean Pit Vipers *Bothrops caribbaeus* and *Bothrops lanceolatus*: Correlation with Toxicological Activities and Immunoreactivity of a Heterologous Antivenom. *Journal of Proteome Research* **2008**, *7*, 4396–4408, doi:10.1021/pr8003826.
113. Savage, J.M. *The Amphibians and Reptiles of Costa Rica. A Herpetofauna between Two Continents, between Two Seas*; The University of Chicago: Chicago and London, **2002**.
114. Lamar, W.; Sasa, Mahmood. A New Species of Hognose Pitviper, Genus *Porthidium*, from the Southwestern Pacific of Costa Rica (Serpentes: Viperidae). *Revista de Biología Tropical*. **2003**, *51*, 797–804.
115. Bocourt, M.F. Descriptions de Quelques Crotaliens Nouveaux Appartenant au Genre *Bothrops*, Recueillis Dans Le Guatémala. *Annales des sciences naturelles., Paris* **1868**, 201–202.
116. Schlegel, H. Description d'une Nouvelle Espèce Du Genre *Trigonocephale* (*Trigonocephalus lansbergii*). *Mag. Zool. Rept* **1841**, 1–3.
117. Monteza-Moreno, C.M.; Ramos, C.; Martínez, V.; Sasa, M. On the identity of hog-nosed pit-vipers from western panama: a review of specimens of *Porthidium lansbergii* (Schlegel, 1841) in lower Central America. *Tecnociencia (Panama)* **2020**, *22*, 27–44, doi:10.48204/j.tecno.v22n2a2.
118. De arco-Rodríguez, B.; Montealegre-Sánchez, L.; Solano-Redondo, L.; Castro-Herrera, F.; Ortega, J.G.; Castillo, A.; Vargas-Zapata, C.; Jiménez-Charris, E. Phylogeny and Toxicological Assessments of Two *Porthidium lansbergii lansbergii* Morphotypes from the Caribbean Region of Colombia. *Toxicon* **2019**, *166*, 56–65, doi:10.1016/j.toxicon.2019.05.010.

119. Cisneros-Heredia, D.F.; Yáñez-Muñoz, M. Reptilia, Viperidae, Crotalinae, *Porthidium nasutum*: Distribution Extension and Remarks on Its Range and Records. *Check List* **2005**, *1*, 16–17, doi:10.15560/1.1.16.
120. Warrell, D.A. Snakebites in Central and South America: Epidemiology, Clinical Features, and Clinical Management. In *Venomous Reptiles of the Western Hemisphere*; Campbell, J.A., Lamar, W.W., Comstock, Eds.; Cornell University: Ithaca, NY, **2004**; Vol. 2, pp. 709–761.
121. Reyes-Velasco, J.; Cox, C.L.; Jones, J.M.; Borja, M.; Campbell, J.A. how many species of rattlesnakes are there in the *Crotalus durissus* species group (Serpentes: Crotalidae); *Revista Latinoamericana de Herpetologia* **2022**, *5*, 43–55, doi:10.22201/fc.25942158e.2022.1.330.
122. Humboldt, A. Sur Deux Nouvelles Espèces de Crotles. In: Humboldt and Bonpland, 1813. *Recueil d'observations de zoologie*. **1811**, *2*, 1–8.
123. Hoge, A.R. Preliminary Account on Neotropical Crotalinae (Serpentes Viperidae). *Mem Inst Butantan* **1966**, *32*, 137–184.
124. García-Perez, J. Una Nueva Especie de Cascabel (Serpentes: Crotalidae) Para El Bolsón Árido de Lagunillas, Cordillera de Mérida, Venezuela. *Latin American scientific contribution to ecology* **1995**, *3*, 7–12.
125. Harris, H.S.JR.; Simmons, R.S. A New Subspecies of *Crotalus durissus* (Serpentes: Crotalidae) from the Rupununi Savanna of Southwestern Guyana. *Instituto Butantan* **1978**, 305–311.
126. van Lidth de Jeude, T.W. On a Collection of Reptiles and Fishes from the West-Indies. *Notes from the Leyden Museum* **1887**, *9*, 129–139.
127. Klauber, Laurence. A New Species of Rattlesnake from Venezuela. *Transactions of the San Diego Society of Natural History* **1941**, *9*, 333–336.
128. Wallach, Van.; Williams, K.; Boundy, J. Snakes of the World: A Catalogue of Living and Extinct Species. *Taylor and Francis, CRC Press* **2014**, 1–1237.
129. Zaher, H.; Murphy, R.W.; Arredondo, J.C.; Graboski, R.; Machado-Filho, P.R.; Mahlow, K.; Montingelli, G.G.; Quadros, A.B.; Orlov, N.L.; Wilkinson, M.; et al. Large-Scale Molecular Phylogeny, Morphology, Divergence-Time Estimation, and the Fossil Record of Advanced Caenophidian Snakes (Squamata: Serpentes). *PLoS One* **2019**, *14*, e0216148, doi:10.1371/journal.pone.0216148.
130. Sawaya, R.J.; M.O.A.V. & M.M. Composition and Natural History of a Cerrado Snake Assemblage at Itirapina, São Paulo State, Southeastern Brazil. *Biota Neotropica* **2008**, *8*, 127–149.
131. Hartmann, P.A. Ecology of a Snake Assemblage in the Atlantic Forest of Southeastern Brazil. *Papéis Avulsos de Zoologia* **2009**, *49*, 343–360.
132. McDiarmid, R.W.; Campbell, J.A.; Touré, T.A. *Snake Species of the World: A Taxonomic and Geographic Reference*; 1st ed.; *Herpetologists' League*, **1999**; Vol. 1.
133. Fernandes, D.S.; Franco, F.L.; Fernandes, R. Systematic revision of the genus *Lachesis* Daudin, 1803 (Serpentes, Viperidae). *Herpetologica* **2004**, *60*, 245–260, doi:10.1655/02-85.
134. Cubides-Cubillos, Sergio.; Loaiza-López, F.; Molina-Betancourth, J.; Agudelo-Sánchez, W. *Lachesis acrochorda* (García, 1896). *Catálogo de Anfibios y Reptiles de Colombia* **2021**, *7*, 39–50.

135. Barrio-Amorós, C.L.; Corrales, G.; Rodríguez, S.; Culebras, J.; Dwyer, Q.; Flores, D.A. The Bushmasters (*Lachesis* Spp.): Queens of the Rainforest. *Reptiles & Amphibians* **2020**, *27*, 358–381, doi:10.17161/randa.v27i3.14978.
136. Madrigal, M.; Sanz, L.; Flores-Díaz, M.; Sasa, M.; Núñez, V.; Alape-Girón, A.; Calvete, J.J. Snake Venomics across Genus *Lachesis*. Ontogenetic Changes in the Venom Composition of *Lachesis stenophrys* and Comparative Proteomics of the Venoms of Adult *Lachesis melanocephala* and *Lachesis acrochorda*. *Journal in Proteomics* **2012**, *77*, 280–297, doi:10.1016/j.jprot.2012.09.003.
137. Diniz-Sousa, R.; Moraes, J. do N.; Rodrigues-da-Silva, T.M.; Oliveira, C.S.; Caldeira, C.A. A Brief Review on the Natural History, Venomics and the Medical Importance of Bushmaster (*Lachesis*) Pit Viper Snakes. *Toxicon X* **2020**, *7*, 100053, doi:10.1016/j.toxcx.2020.100053.
138. Díaz-Ricaurte, Juan.; Guevara-Molina, C.; Cubides-Cubillos, Sergio. *Lachesis muta* (Linnaeus 1766). *Catálogo de Anfibios y Reptiles de Colombia* **2017**, *3*, 20–24.
139. Arteaga, A.; Pyron, R.A.; Batista, A.; Vieira, J.; Meneses Pelayo, E.; Smith, E.N.; Barrio Amorós, C.L.; Koch, C.; Agne, S.; Valencia, J.H.; et al. Systematic Revision of the Eyelash Palm-Pitviper *Bothriechis schlegelii* (Serpentes, Viperidae), with the Description of Five New Species and Revalidation of Three. *Journal of Systematics and Evolution* **2024**, *8*, 15–64, doi:10.3897/evolsyst.8.114527.
140. Calvete, J.J.; Escolano, J.; Sanz, L. Snake Venomics of *Bitis* Species Reveals Large Intragenus Venom Toxin Composition Variation: Application to Taxonomy of Congeneric Taxa. *Journal of Proteome Research* **2007**, *6*, 2732–2745, doi:10.1021/pr0701714.
141. Fry, B.G. From Genome to “Venome”: Molecular Origin and Evolution of the Snake Venom Proteome Inferred from Phylogenetic Analysis of Toxin Sequences and Related Body Proteins. *Genome Research* **2005**, *15*, 403–420, doi:10.1101/gr.3228405.
142. Earl, S.T.H.; Birrell, G.W.; Wallis, T.P.; St Pierre, L.D.; Masci, P.P.; de Jersey, J.; Gorman, J.J.; Lavin, M.F. Post-Translational Modification Accounts for the Presence of Varied Forms of Nerve Growth Factor in Australian Elapid Snake Venoms. *Proteomics* **2006**, *6*, 6554–6565, doi:10.1002/pmic.200600263.
143. Chippaux, J. P.; Williams, V.; White, J.; Chippaux, J. P.; Williabis, V. State Toxinology Service, Adelaide Children’s Hospital; **1991**; Vol. 29.
144. Casewell, N.R.; Jackson, T.N.W.; Laustsen, A.H.; Sunagar, K. Causes and Consequences of Snake Venom Variation. *Trends in Pharmacological Sciences* **2020**, *41*, doi:10.1016/j.tips.2020.05.006.
145. Fry, B.G.; Wüster, W. Assembling an Arsenal: Origin and Evolution of the Snake Venom Proteome Inferred from Phylogenetic Analysis of Toxin Sequences. *Molecular Biology and Evolution* **2004**, *21*, 870–883, doi:10.1093/molbev/msh091.
146. Wüster, W.; Peppin, L.; Pook, C.E.; Walker, D.E. A Nesting of Vipers: Phylogeny and Historical Biogeography of the Viperidae (Squamata: Serpentes). *Molecular Phylogenetics and Evolution* **2008**, *49*, doi:10.1016/j.ympev.2008.08.019.
147. Fry, B.G.; Winkel, K.D.; Wickramaratna, J.C.; Hodgson, W.C.; Wüster, W. Effectiveness of Snake Antivenom: Species and Regional Venom

- Variation and Its Clinical Impact. *Journal of Toxicology: Toxin Reviews* **2003**, 22, 23–34, doi:10.1081/TXR-120019018.
148. Calvete, J.J.; Borges, A.; Segura, Á.; Flores-Díaz, M.; Alape-Girón, A.; Gutiérrez, J.M.; Diez, N.; De Sousa, L.; Kiriakos, D.; Sánchez, E.; et al. Snake Venomics and Antivenomics of *Bothrops colombiensis*, a Medically Important Pitviper of the *Bothrops atrox-asper* Complex Endemic to Venezuela: Contributing to Its Taxonomy and Snakebite Management. *Journal in Proteomics* **2009**, 72, 227–240, doi:10.1016/j.jprot.2009.01.005.
149. Klupczynska, A.; Pawlak, M.; Kokot, Z.J.; Matysiak, J. Application of Metabolomic Tools for Studying Low Molecular-Weight Fraction of Animal Venoms and Poisons. *Toxins (Basel)* **2018**, 10, 306, <https://doi.org/10.3390/toxins10080306>
150. Modahl, C.M.; Brahma, R.K.; Koh, C.Y.; Shioi, N.; Kini, R.M. Omics Technologies for Profiling Toxin Diversity and Evolution in Snake Venom: Impacts on the Discovery of Therapeutic and Diagnostic Agents. *Annual Review of Animal Bioscience* **2020**, 8, 91–116.
151. Abd El-Aziz, T.M.; Soares, A.G.; Stockand, J.D. Advances in Venomics: Modern Separation Techniques and Mass Spectrometry. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences* **2020**, 1160.
152. Walker, A.A.; Robinson, S.D.; Hamilton, B.F.; Undheim, E.A.B.; King, G.F. Deadly Proteomes: A Practical Guide to Proteotranscriptomics of Animal Venoms. *Proteomics* **2020**, 20, doi:10.1002/pmic.201900324.
153. Damm, M.; Hempel, B.F.; Süssmuth, R.D. Old World Vipers—a Review about Snake Venom Proteomics of Viperinae and Their Variations. *Toxins (Basel)* **2021**, 13.
154. Chippaux, J.-P.; Williams, V.; White, J. Snake Venom Variability: Methods of Study, Results and Interpretation. *Toxicon* **1991**, 29, doi:10.1016/0041-0101(91)90116-9.
155. Arbuckle, K. From Molecules to Macroevolution: Venom as a Model System for Evolutionary Biology across Levels of Life. *Toxicon X* **2020**, 6, doi:10.1016/j.toxcx.2020.100034.
156. Zancolli, G.; Casewell, N.R. Venom Systems as Models for Studying the Origin and Regulation of Evolutionary Novelty. *Molecular Biology and Evolution* **2020**, 37, doi:10.1093/molbev/msaa133.
157. Fry, B.G. *Venomous Reptiles and Their Toxins. Evolution, Pathophysiology, and Biodiscovery*; Oxford University Press, USA., Ed.; Oxford University Press: New York, NY., **2015**; Vol. 1.
158. Mackessy, S.P.; Saviola, A.J. Understanding Biological Roles of Venoms Among the Caenophidia: The Importance of Rear-Fanged Snakes. *Integrative and Comparative Biology* **2016**, 56, 1004–1021, doi:10.1093/icb/icw110.
159. Fry, B.G.; Scheib, H.; van der Weerd, L.; Young, B.; McNaughtan, J.; Ryan Ramjan, S.F.; Vidal, N.; Poelmann, R.E.; Norman, J.A. Evolution of an Arsenal: Structural and Functional Diversification of the Venom System in the Advanced Snakes (Caenophidia). *Molecular and Cellular Proteomics* **2008**, 7, 215–246, doi:10.1074/mcp.M700094-MCP200.
160. Jackson, T.; Koludarov, I.; Ali, S.; Dobson, J.; Zdenek, C.; Dashevsky, D.; op den Brouw, B.; Masci, P.; Nouwens, A.; Josh, P.; et al. Rapid Radiations and the Race to Redundancy: An Investigation of the Evo-

- lution of Australian Elapid Snake Venoms. *Toxins (Basel)* **2016**, *8*, 309, doi:10.3390/toxins8110309.
161. Casewell, N.R.; Wüster, W.; Vonk, F.J.; Harrison, R.A.; Fry, B.G. Complex Cocktails: The Evolutionary Novelty of Venoms. *Trends in Ecology & Evolution* **2013**, *28*, 219–229, doi:10.1016/j.tree.2012.10.020.
 162. Giorgianni, M.W.; Dowell, N.L.; Griffin, S.; Kassner, V.A.; Selegue, J.E.; Carroll, S.B. The Origin and Diversification of a Novel Protein Family in Venomous Snakes. *Proceedings of the National Academy of Sciences* **2020**, *117*, 10911–10920, doi:10.1073/pnas.1920011117.
 163. Amorim, F.; Costa, T.; Baiwir, D.; De Pauw, E.; Quinton, L.; Sampaio, S. Proteo-peptidomic, Functional and Immunoreactivity Characterization of *Bothrops moojeni* Snake Venom: Influence of Snake Gender on Venom Composition. *Toxins (Basel)* **2018**, *10*, 177, doi:10.3390/toxins10050177.
 164. Queiroz, G.P.; Pessoa, L.A.; Portaro, F.C.V.; Furtado, M. de F.D.; Tambourgi, D. V. Interspecific Variation in Venom Composition and Toxicity of Brazilian Snakes from *Bothrops* Genus. *Toxicon* **2008**, *52*, 842–851, doi:10.1016/j.toxicon.2008.10.002.
 165. Kazandjian, T.D.; Petras, D.; Robinson, S.D.; van Thiel, J.; Greene, H.W.; Arbuckle, K.; Barlow, A.; Carter, D.A.; Wouters, R.M.; Whiteley, G.; et al. Convergent Evolution of Pain-Inducing Defensive Venom Components in Spitting Cobras. *Science* **2021**, *371*, 386–390, doi:10.1126/science.abb9303.
 166. Barua, A.; Mikheyev, A.S. Toxin Expression in Snake Venom Evolves Rapidly with Constant Shifts in Evolutionary Rates. *Proceedings of the Royal Society: Biological Sciences* **2020**, *287*, 20200613, doi:10.1098/rspb.2020.0613.
 167. Cubides-Cubillos, S.D.; Alarcón-Pérez, J.C. Accidente Ofídico En Antioquia, Colombia: Análisis Etnobiológico de Las Construcciones Culturales. *Revista Etnobiología* **2018**, *16*, 18–29.
 168. Chippaux, J.-P. Snakebite Envenomation Turns Again into a Neglected Tropical Disease! *Journal of Venomous Animals and Toxins including Tropical Diseases* **2017**, *23*, 38, doi:10.1186/s40409-017-0127-6.
 169. Chippaux, J.-P. Epidemiology of Envenomations by Terrestrial Venomous Animals in Brazil Based on Case Reporting: From Obvious Facts to Contingencies. *Journal of Venomous Animals and Toxins including Tropical Diseases* **2015**, *21*, 13, doi:10.1186/s40409-015-0011-1.
 170. Boldrini-França, J.; Corrêa-Netto, C.; Silva, M.M.S.; Rodrigues, R.S.; De La Torre, P.; Pérez, A.; Soares, A.M.; Zingali, R.B.; Nogueira, R.A.; Rodrigues, V.M.; et al. Snake Venomics and Antivenomics of *Crotalus durissus* Subspecies from Brazil: Assessment of Geographic Variation and Its Implication on Snakebite Management. *Journal in Proteomics* **2010**, *73*, 1758–1776, doi:10.1016/j.jprot.2010.06.001.
 171. Rodríguez-Vargas, A.; Vega, N.; Reyes-Montaño, E.; Corzo, G.; Neri-Castro, E.; Clement, H.; Ruiz-Gómez, F. Intraspecific Differences in the Venom of *Crotalus durissus cumanensis* from Colombia. *Toxins (Basel)* **2022**, *14*, doi:10.3390/toxins14080532.
 172. Talukdar, A.; Maddhesiya, P.; Namsa, N.D.; Doley, R. Snake Venom Toxins Targeting the Central Nervous System. *Toxin Reviews* **2023**, *42*, 382–406, doi:10.1080/15569543.2022.2084418.

173. Lewis, R.; Gutmann, L. Snake Venoms and the Neuromuscular Junction. *Semin Neurol* **2004**, *24*, 175–179, doi:10.1055/s-2004-830904.
174. Rossetto, O.; Morbiato, L.; Caccin, P.; Rigoni, M.; Montecucco, C. Presynaptic Enzymatic Neurotoxins. *Journal Neurochem* **2006**, *97*, 1534–1545, doi:10.1111/j.1471-4159.2006.03965.x.
175. Gawade, S.P. Snake Venom Neurotoxins: Pharmacological Classification. *Journal of Toxicology: Toxin Reviews* **2004**, *23*, 37–96, doi:10.1081/TXR-120030647.
176. Calvete, J.J.; Sanz, L.; Cid, P.; De La Torre, P.; Flores-Díaz, M.; Dos Santos, M.C.; Borges, A.; Bremo, A.; Angulo, Y.; Lomonte, B.; et al. Snake Venomics of the Central American Rattlesnake *Crotalus simus* and the South American *Crotalus durissus* Complex Points to Neurotoxicity as an Adaptive Paedomorphic Trend along *Crotalus* Dispersal in South America. *Journal of Proteome Research* **2010**, *9*, 528–544, doi:10.1021/pr9008749.
177. Batista da Cunha, D.; Pupo Silvestrini, A.V.; Gomes da Silva, A.C.; Maria de Paula Estevam, D.; Pollettini, F.L.; de Oliveira Navarro, J.; Alves, A.A.; Remédio Zeni Beretta, A.L.; Annichino Bizzacchi, J.M.; Pereira, L.C.; et al. Mechanistic Insights into Functional Characteristics of Native Crotoamine. *Toxicon* **2018**, *146*, 1–12, doi:10.1016/j.toxicon.2018.03.007.
178. Rádis-Baptista, G.; Kerkis, I. Crotoamine, a Small Basic Polypeptide Myo-toxin from Rattlesnake Venom with Cell-Penetrating Properties. *Curr Pharm Des* **2011**, *17*(38), 4351–4361, doi:10.2174/138161211798999429
179. Schenberg, S. Geographical Pattern of Crotoamine Distribution in the Same Rattlesnake Subspecies. *Science* (1979) **1959**, *129*, 1361–1363.
180. Otero, R.; Osorio, R.G.; Valderrama, R.; Giraldo, C.A. Efectos farmacológicos y enzimáticos de los venenos de serpientes de Antioquia y Chocó (Colombia); *Toxicon* **1992**, *30* (5-6): 611–620, doi: [https://doi.org/10.1016/0041-0101\(92\)90855-Y](https://doi.org/10.1016/0041-0101(92)90855-Y)
181. Mora-Obando, D.; Plaid, D.; Lomonte, B.; Guerrero-Vargas, J.A.; Ayerbe, S.; Calvete, J.J. Antivenomics and in Vivo Preclinical Efficacy of Six Latin American Antivenoms towards Southwestern Colombian *Bothrops asper* Lineage Venoms. *PLOS Neglected Tropical Diseases* **2021**, *15*, 1–36, doi:10.1371/journal.pntd.0009073.
182. Arévalo-Páez, M.; Rada-Vargas, E.; Betancur-Hurtado, C.; Renjifo, J.M.; Renjifo-Ibáñez, C. Neuromuscular Effect of Venoms from Adults and Juveniles of *Crotalus durissus cumanensis* (Humboldt, 1811) from Guajira, Colombia. *Toxicon* **2017**, *139*, 41–44, doi:10.1016/j.toxicon.2017.09.016.
183. Pereañez, J.A.; Núñez, V.; Huancahuire-Vega, S.; Marangoni, S.; Ponce-Soto, L.A. Biochemical and Biological Characterization of a PLA₂ from Crotoxin Complex of *Crotalus durissus cumanensis*. *Toxicon* **2009**, *53*, 534–542, doi:10.1016/j.toxicon.2009.01.021.
184. Mora-Obando, D.; Salazar-Valenzuela, D.; Pla, D.; Lomonte, B.; Guerrero-Vargas, J.A.; Ayerbe, S.; Gibbs, H.L.; Calvete, J.J. Venom Variation in *Bothrops asper* Lineages from North-Western South America. *Journal in Proteomics* **2020**, *229*, 103945, doi:10.1016/j.jprot.2020.103945.
185. Jiménez-Charris, E.; Montoya-Gómez, A.; Torres, J.K.; Gómez-Díaz, M.; Bolívar-García, W. First Functional and Proteomic Analysis of *Bothrops asper* Snake Venom from Gorgona Island - Colombia, and Its Compa-

- native Characterization with Two Colombian Southwest Ecoregions. *Biochimie* **2022**, *194*, 19–27, doi:10.1016/j.biochi.2021.12.005.
186. Quintana, J.; Otero, R.; Nuñez, V.; Toro, F. Estudio de La Variabilidad En El Veneno de 2 Poblaciones de *Bothriechis schlegelii* Del Suroeste y Norte de Antioquia y Correlación Morfométrica. *Iatreia* **2000**, 1–107.
 187. Montealegre-Sánchez, L.; Montoya-Gómez, A.; Jiménez-Charris, E. Individual Variations in the Protein Profiles and Functional Activities of the Eyelash Palm Pit-Viper (*Bothriechis schlegelii*) Venom from the Colombian Southwest Region. *Acta Tropica* **2021**, *223*, 106113, doi:10.1016/j.actatropica.2021.106113.
 188. Lomonte, B.; Escolano, J.; Fernández, J.; Sanz, L.; Angulo, Y.; Gutiérrez, J.M.; Calvete, J.J. Snake Venomics and Antivenomics of the Arboreal Neotropical Pitvipers *Bothriechis lateralis* and *Bothriechis schlegelii*. *Journal of Proteome Research* **2008**, *7*, 2445–2457, doi:10.1021/pr8000139.
 189. Gutiérrez, J.; Escalante, T.; Rucavado, A.; Herrera, C. Hemorrhage Caused by Snake Venom Metalloproteinases: A Journey of Discovery and Understanding. *Toxins (Basel)* **2016**, *8*, 93, doi:10.3390/toxins8040093.
 190. Otero, R.; Gutiérrez, J.; Beatriz Mesa, M.; Duque, E.; Rodríguez, O.; Luis Arango, J.; Gómez, F.; Toro, A.; Cano, F.; María Rodríguez, L.; et al. Complications of *Bothrops*, *Porthidium*, and *Bothriechis* Snakebites in Colombia. A Clinical and Epidemiological Study of 39 Cases Attended in a University Hospital. *Toxicon* **2002**, *40*, 1107–1114, doi:10.1016/S0041-0101(02)00104-6.
 191. Otero-Patiño, R. Snake Bites in Colombia. In: Vogel, CW., Seifert, S., Tambourgi, D. (eds) *Clinical Toxinology in Australia, Europe, and Americas*. *Toxinology*. Springer, Dordrecht; **2018**; pp. 3–50. https://doi.org/10.1007/978-94-017-7438-3_41
 192. Preciado, L.M.; Pereañez, J.A.; Comer, J. Potential of Matrix Metalloproteinase Inhibitors for the Treatment of Local Tissue Damage Induced by a Type P-I Snake Venom Metalloproteinase. *Toxins (Basel)* **2019**, *12*, 8, doi:10.3390/toxins12010008.
 193. Pereañez, J.A.; Preciado, L.M.; Fernández, J.; Camacho, E.; Lomonte, B.; Castro, F.; Cañas, C.A.; Galvis, C.; Castaño, S. Snake Venomics, Experimental Toxic Activities and Clinical Characteristics of Human Envenomation by *Bothrocophias myersi* (Serpentes: Viperidae) from Colombia. *Journal in Proteomics* **2020**, *220*, doi:10.1016/j.jprot.2020.103758.
 194. Sevilla-Sánchez, M.-J.; Guerrero-Vargas, J.A.; Ayerbe-González, S.; Calderón-Leytón, J.J.; Lomonte, B.; Mora-Obando, D. Toxinological Profile and Histopathological Alterations Induced by *Bothrocophias campbelli* Venom from Colombia. *Acta Tropica* **2024**, *250*, 107094, doi:10.1016/j.actatropica.2023.107094.
 195. Jiménez-Charris, E.; Montealegre-Sanchez, L.; Solano-Redondo, L.; Mora-Obando, D.; Camacho, E.; Castro-Herrera, F.; Fierro-Pérez, L.; Lomonte, B. Proteomic and Functional Analyses of the Venom of *Porthidium lansbergii lansbergii* (Lansberg's Hognose Viper) from the Atlantic Department of Colombia. *J Proteomics* **2015**, *114*, 287–299, doi:10.1016/j.jprot.2014.11.016.

196. Otero-Patiño, R.; Segura, Á.; Herrera, M.; Angulo, Y.; León, G.; Gutiérrez, J.M.; Barona, J.; Estrada, S.; Pereañez, A.; Quintana, J.C.; et al. Comparative Study of the Efficacy and Safety of Two Polyvalent, Caprylic Acid Fractionated [IgG and F(Ab')₂] Antivenoms, in *Bothrops asper* Bites in Colombia. *Toxicon* **2012**, *59*, 344–355, doi:10.1016/j.toxicon.2011.11.017.
197. Kalil, J.; Fan, H.W. Production and Utilization of Snake Antivenoms in South America. In; **2017**; pp. 81–101.
198. Bermúdez-Méndez, E.; Fuglsang-Madsen, A.; Føns, S.; Lomonte, B.; Gutiérrez, J.; Laustsen, A. Innovative Immunization Strategies for Antivenom Development. *Toxins (Basel)* **2018**, *10*, 452, doi:10.3390/toxins10110452.
199. Gutiérrez, J.M.; León, G.; Burnouf, T. Antivenoms for the Treatment of Snakebite Envenomings: The Road Ahead. *Biologicals* **2011**, *39*, 129–142.
200. Theakston, R.D.G.; Reid, H.A. Development of Simple Standard Assay Procedures for the Characterization of Snake Venoms. *Bulletin of the World Health Organization* **1983**, *61*, 949–956.
201. Rojas, G.; Jiménez, J.M.; Gutiérrez, J.M. Caprylic Acid Fractionation of Hyperimmune Horse Plasma: Description of a Simple Procedure for Antivenom Production; *Toxicon* **1994**, *32*, 351–363, [https://doi.org/10.1016/0041-0101\(94\)90087-6](https://doi.org/10.1016/0041-0101(94)90087-6).
202. Laloo, D.G.; Theakston, R.D.G. Snake Antivenoms. *Journal of Toxicology: Clinical Toxicology* **2003**, *41*, 277–290, doi:10.1081/CLT-120021113.
203. Bogarín, G.; Romero, M.; Rojas, G.; Lutsch, C.; Casadamont, M.; Lang, J.; Otero, R.; Gutiérrez, J.M. Neutralization, by a Monospecific *Bothrops lanceolatus* Antivenom, of Toxic Activities Induced by Homologous and Heterologous *Bothrops* Snake Venoms. *Toxicon* **1999**, *37*, 551–557, doi:10.1016/S0041-0101(98)00193-7.
204. Bucher, B.; Canongel, D.; Thomas, L.; Tyburn, B. Clinical Indicators of Envenoming and Serum Levels of Venom Antigens Bitten by *Bothrops lanceolatus* in Martinique. *Transactions of The Royal Society of Tropical Medicine*. **1997** (2):186–90. doi: 10.1016/s0035-9203(97)90219-4..
205. Gutiérrez, J.M.; Theakston, R.D.G.; Warrell, D.A. Confronting the Neglected Problem of Snake Bite Envenoming: The Need for a Global Partnership. *PLoS Medicine* **2006**, *3*, 0727–0731.
206. Layfield, H.J.; Williams, H.F.; Ravishankar, D.; Mehmi, A.; Sonavane, M.; Salim, A.; Vaiyapuri, R.; Lakshminarayanan, K.; Vallance, T.M.; Bicknell, A.B.; et al. Repurposing Cancer Drugs Batimastat and Marimastat to Inhibit the Activity of a Group I Metalloprotease from the Venom of the Western Diamondback Rattlesnake, *Crotalus atrox*. *Toxins (Basel)* **2020**, *12*, 309, doi:10.3390/toxins12050309.
207. Calvete, J.J.; Pérez, A.; Lomonte, B.; Sánchez, E.E.; Sanz, L. Snake Venomics of *Crotalus tigris*: The Minimalist Toxin Arsenal of the Deadliest Neartic Rattlesnake Venom. Evolutionary Clues for Generating a Pan-Specific Antivenom against Crotalid Type II Venoms. *Journal of Proteome Research* **2012**, *11*, 1382–1390, doi:10.1021/pr201021d.
208. Calvete, J.J.; Fasoli, E.; Sanz, L.; Boschetti, E.; Righetti, P.G. Exploring the Venom Proteome of the Western Diamondback Rattlesnake, *Crotalus atrox*, via Snake Venomics and Combinatorial Peptide Ligand Library Approaches. *Journal of Proteome Research* **2009**, *8*, 3055–3067, doi:10.1021/pr900249q.

209. Romero-Giraldo, L.E.; Pulido, S.; Berrío, M.A.; Flórez, M.F.; Rey-Suárez, P.; Nuñez, V.; Pereañez, J.A. Heterologous Expression and Immunogenic Potential of the Most Abundant Phospholipase A2 from Coral Snake *Micrurus dumerilii* to Develop Antivenoms. *Toxins (Basel)* **2022**, *14*, 825, doi:10.3390/toxins14120825.
210. Carvalho, B.M.A.; Santos, J.D.L.; Xavier, B.M.; Almeida, J.R.; Resende, L.M.; Martins, W.; Marcussi, S.; Marangoni, S.; Stábeli, R.G.; Calderon, L.A.; et al. Snake Venom PLA₂s Inhibitors Isolated from Brazilian Plants: Synthetic and Natural Molecules. *BioMed Research International* **2013**, *2013*, 1–8, doi:10.1155/2013/153045.
211. Gómez-Betancur, I.; Gogineni, V.; Salazar-Ospina, A.; León, F. Perspective on the Therapeutics of Anti-Snake Venom. *Molecules* **2019**, *24*, 3276, doi:10.3390/molecules24183276.
212. Quiroz, S.; Henao Castañeda, I.C.; Granados, J.; Patiño, A.C.; Preciado, L.M.; Pereañez, J.A. Inhibitory Effects of Varespladib, CP471474, and Their Potential Synergistic Activity on *Bothrops asper* and *Crotalus durissus cumanensis* Venoms. *Molecules* **2022**, *27*, 8588, doi:10.3390/molecules27238588.
213. Mora-Obando, D.; Lomonte, B.; Pla, D.; Guerrero-Vargas, J.A.; Ayerbe-González, S.; Gutiérrez, J.M.; Sasa, M.; Calvete, J.J. Half a Century of Research on *Bothrops asper* Venom Variation: Biological and Biomedical Implications. *Toxicon* **2023**, *221*, 106983, doi:10.1016/j.toxicon.2022.106983.
214. Calvete, J.J.; Rodríguez, Y.; Quesada-Bernat, S.; Pla, D. Toxin-Resolved Antivenomics-Guided Assessment of the Immunorecognition Landscape of Antivenoms. *Toxicon* **2018**, *148*, 107–122, doi:10.1016/j.toxicon.2018.04.015.
215. Nguyen, L.-T.; Schmidt, H.A.; von Haeseler, A.; Minh, B.Q. IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Molecular Biology and Evolution* **2015**, *32*, 268–274, doi:10.1093/molbev/msu300.
216. Kalyaanamoorthy, S.; Minh, B.Q.; Wong, T.K.F.; von Haeseler, A.; Jermini, L.S. ModelFinder: Fast Model Selection for Accurate Phylogenetic Estimates. *Nature Methods* **2017**, *14*, 587–589, doi:10.1038/nmeth.4285.
217. Revell, L.J. Phytools: An R Package for Phylogenetic Comparative Biology (and Other Things). *Methods in Ecology and Evolution* **2012**, *3*, 217–223, doi:10.1111/j.2041-210X.2011.00169.x.
218. Warrell, D.A. Snake Bite. *The Lancet* **2010**, *375*, 77–88.
219. Ramakrishnan, M.A. Determination of 50% Endpoint Titer Using a Simple Formula. *World Journal of Virology* **2016**, *5*, 85, doi:10.5501/wjv.v5.i2.85.
220. Sells, P.G. Animal Experimentation in Snake Venom Research and in Vitro Alternatives. *Toxicon* **2003**, *42*, 115–133.
221. Boivin, G.P.; Hickman, D.L.; Creamer-Hente, M.A.; Pritchett-Corning, K.R.; Bratcher, N.A. Review of CO₂ as a Euthanasia Agent for Laboratory Rats and Mice. *Journal of the American Association for Laboratory Animal Science* **2017**, *56*, 491–499.
222. Reyes-Velasco, J. A revision of recent taxonomic changes to the eyelash palm pitviper (Serpentes, Viperidae, *Bothriechis schlegelii*). *Herpetozoa* **2024**, *37*, 1–14, doi:10.3897/herpetozoa.